

Master Thesis

Soil Carbon Dioxide Fluxes from Two Austrian Forests under Drying Rewetting Stress

submitted by

Franco Alexis GONZÁLEZ

in the framework of the Master programme

International Master in Soils and Global Change (IMSOGLO)

in partial fulfilment of the requirements for the academic degree

Master of Science

Vienna, August 2022

Co-supervisor:
Univ. Prof. Dr Alexander Knohl
Centre of Biodiversity and Sustainable Land Use (CBL)
Department of Bioclimatology,
Faculty of Forest Sciences,
Georg-August-Universität, Göttingen, Germany

Supervisor:
Univ. Prof. DI Dr. Sophie Zechmeister-Boltenstern
Institute of Soil Research
Department of Forest and Soil Sciences,
University of Natural Resources and Life Sciences,
Vienna, Austria



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Main Supervisor:

Univ. Prof. Dr Phil. Sophie Zechmeister-Boltenstern

Institute of Soil Research

Department of Forest and Soil Sciences

University of Natural Resources and Life Sciences Vienna

Co-Supervisors:

Dr Eugenio Diaz-Pines

Institute of Soil Research

Department of Forest and Soil Sciences.

University of Natural Resources and Life Sciences Vienna

MSc. Dylan Goff

Institute of Soil Research

Department of Forest and Soil Sciences.

University of Natural Resources and Life Sciences Vienna

Univ. Prof. Dr Alexander Knohl

Centre of Biodiversity and Sustainable Land Use (CBL)

Department of Bioclimatology

Faculty of Forest Sciences

Georg-August-Universität Göttingen

Affidavit

I hereby declare that I have authored this master thesis independently, and that I have not used any assistance other than that which is permitted. The work contained herein is my own except where explicitly stated otherwise. All ideas taken in wording or in basic content from unpublished sources or from published literature are duly identified and cited, and the precise references included.

I further declare that this master thesis has not been submitted, in whole or in part, in the same or a similar form, to any other educational institution as part of the requirements for an academic degree.

I hereby confirm that I am familiar with the standards of Scientific Integrity and with the guidelines of Good Scientific Practice, and that this work fully complies with these standards and guidelines.

Vienna, August 15th

Franco GONZÁLEZ (*manu propria*)

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Abstract

More frequent and severe extreme weather and climate events are among the outcomes of global warming. Hence, more frequent and intense dry-rewetting (DRW) stress is expected. Soils play an essential role in controlling GHGs' atmospheric concentration, not only by direct emissions after the turnover of elements but also by sequestering vast amounts of C. Soil organic carbon (SOC) is the largest terrestrial C pool, estimated at around 1700 Pg C, roughly thrice and twice as big as the vegetation and the atmospheric pool, respectively. R_s , performed by microorganisms and plant roots, releases vast amounts of CO_2 ($\sim 94 \text{ Pg C yr}^{-1}$), representing the largest C flux between land and the atmosphere after photosynthesis and being almost ten times higher than fossil fuel emissions on an annual basis. Soil temperature and moisture are primarily known to control R_s temporal variability, although other soil parameters that regulate microbial activity can also be important when spatial variability is considered. A prolonged period without water input dries the soil below the field capacity, modifying several physical and biological processes. Rewetting dry soils usually triggers a disproportionated burst of CO_2 , a phenomenon known as the "Birch effect", that would be decoupled from microbial growth. Twentieth-century industrial fixation of N_2 into reactive N (Nr) forms for posterior release in ecosystems resulted in a "cascade" of intended and unintended consequences for the C cycle still unclear. Among several effects, the increased Nr over temperate forests has been generally described as reducing R_s and increasing C sequestration. This study aims to assess the effect of drying-rewetting stress on the soil CO_2 fluxes and to uncover the controlling soil parameters in one mature European beech (*Fagus sylvatica* L.) forest in Eastern Austria during one drying-rewetting stress event, under different nitrogen deposition scenarios.

Kurzfassung

Häufigere und heftigere extreme Wetter- und Klimaereignisse gehören zu den Folgen der globalen Erwärmung. Daher wird ein häufigerer und intensiverer Dry-Rewetting (DRW)-Stress erwartet. Böden spielen eine wesentliche Rolle bei der Kontrolle der atmosphärischen Konzentration von Treibhausgasen, nicht nur durch direkte Emissionen nach dem Umsatz von Elementen, sondern auch durch die Bindung großer Mengen an C. Der organische Kohlenstoff im Boden (SOC) ist der höchste terrestrische C-Speicher, der auf etwa 1700 Pg C geschätzt wird, ungefähr dreimal und doppelt so groß wie die Vegetation bzw. der atmosphärische Pool. R_s , die von Mikroorganismen und Pflanzenwurzeln durchgeführt wird, setzt riesige Mengen an CO_2 (~94 Pg C pro Jahr) frei, was den größten C-Fluss zwischen Land und Atmosphäre nach der Photosynthese darstellt und fast zehnmal höher ist als die jährlichen Emissionen aus fossilen Brennstoffen. Es ist in erster Linie bekannt, dass Bodentemperatur und Feuchtigkeit die zeitliche Variabilität von R_s steuern, obwohl andere Bodenparameter, die die mikrobielle Aktivität regulieren, auch wichtig sein können, wenn die räumliche Variabilität berücksichtigt wird. Ein längerer Zeitraum ohne Wasserzufuhr trocknet den Boden unterhalb der Feldkapazität aus, wodurch mehrere physikalische und biologische Prozesse modifiziert werden. Die Wiedervernässung trockener Böden löst in der Regel einen überproportionalen CO_2 -Ausstoß aus, ein als „Birkeneffekt“ bekanntes Phänomen, das vom mikrobiellen Wachstum entkoppelt wäre. Die industrielle Fixierung von N_2 in reaktive $N(Nr)$ -Formen im 20. Jahrhundert zur späteren Freisetzung in Ökosystemen führte zu einer „Kaskade“ beabsichtigter und unbeabsichtigter Folgen für den C-Kreislauf, die noch unklar sind. Neben mehreren Effekten wurde die erhöhte Nr über Wäldern der gemäßigten Breiten allgemein als Verringerung von R_s und Erhöhung der C-Sequestrierung beschrieben. Diese Studie zielt darauf ab, die Wirkung von Trocknungs-Wiedervernässungs-Stress auf die CO_2 -Flüsse im Boden zu bewerten und die kontrollierenden Bodenparameter in einem ausgewachsenen Rotbuchenwald (*Fagus sylvatica* L.) in Ostösterreich während eines Trocknungs-Wiedervernässungs-Stressereignisses unter verschiedenen Stickstoffbedingungen aufzudecken Ablagerungsszenarien.

1. Introduction

1.1. Anthropogenic climate warming and its impacts

Anthropic activity is warming the global surface by enhancing the atmospheric greenhouse effect (IPCC, 2013). The greenhouse gas (GHG) concentration sharply augmented over the last two and a half centuries (ca. 1750; IPCC, 2013), which is attributed to the skyrocketed human-derived emissions of the three main gases, carbon dioxide (CO₂), nitrous oxide (N₂O), and methane (CH₄). After water vapour, the main contributor to the greenhouse effect is CO₂, and its atmospheric concentration rose from ~277 ppm from 1750 to ~414 in 2021 (Friedlingstein et al., 2022). Fossil fuel combustion is the largest source, followed by land use and land cover change (LULCC). Humans have burned ~450 Pg C since 1870 (Le Quéré et al., 2018, Friedlingstein et al., 2021). Likewise, LULCC contributes to GHGs emissions directly by disrupting large C pools (e.g., deforestation) or indirectly by reducing the terrestrial ecosystem's sink capacity, such as soil C sequestration. The area of land dedicated to pasture and crops has risen from an area between 7.5 to 9 million Km² in 1750 to 50 million Km² in 2011 (Ciais et al., 2013). Consequently, the resultant total anthropogenic emissions released is 700 ± 75 Pg C for the period 1750-2019, from which fossil fuels contributed $64\% \pm 15\%$, and the remaining arose from LULCC (Arias et al., 2021), and most of these emissions occurred after 1960 (~70%; Friedlingstein et al., 2021). There has been a strong increasing tendency in anthropogenic-derived CO₂ emissions in the last six decades, reaching its highest levels in the decade 2010-2019, with an average annual value at 10.9 ± 0.9 Pg C yr⁻¹, which 46% of it ending at the atmosphere while 31% was retained by terrestrial ecosystems (land-borne fraction; Friedlingstein et al., 2021) and the oceans mainly absorbed the remaining. The increase was exclusively due to fossil fuel emissions which have risen from 3 to ~10 Pg C yr⁻¹ from 1960 to 2021, while LULCC decreased from 1.6 to 0.8 Pg C yr⁻¹ (Friedlingstein et al., 2021). The land C sink is expected to increase in magnitude with higher atmospheric CO₂ levels, but the share of the emissions captured will decrease, and a warmer and dryer climate will slow the growing sink effect (Arias et al., 2021). As a result, the human warming effect is estimated on 1.09 (0.95-1.20) °C since the 1850-1900 baseline, and it will cross the 1.5 °C level soon (2030-2052; Arias et al., 2021) with important already unavoidable negative consequences for earth ecosystems in the present century no matter the course of actions (e.g., emission reduction), and further catastrophic predictions for the next centuries if emissions do not strongly decline.

More frequent and severe extreme weather and climate events are among the outcomes of global warming (Sherwood & Fu, 2014). Since 1950 there has been good observational proof confirming an increase in frequency and heavy precipitation over most land regions (Arias et al., 2021; Hartman et al., 2013). This would lead to less water available since when the water input exceeds the soil retention capacity, it is not incorporated into the soil and flows away as runoff or deep percolation. Similarly, heatwaves and droughts were more frequent in the last century and are expected to increase alongside global warming (Orth et al., 2016); both types of events also deplete soil water availability due to enhanced evapotranspiration. Hence, more frequent and intense dry-rewetting (DRW) stress is expected.

1.2. Land and soil carbon cycle

Soils play an essential role in controlling GHGs' atmospheric concentration, not only by direct emissions after the turnover of elements but also by sequestering vast amounts of C. Terrestrial plant photosynthesis, termed gross primary production (GPP), removes CO₂ from the atmosphere at a rate estimated between 119 and 169 Pg C yr⁻¹ (Anav et al., 2015), which represents roughly one out of eight atmospheric CO₂ molecules (Reich, 2010) and the major land C sink. GPP mainly depends on the vegetation-growing season (Goulden et al., 1996). Although most of this C is reemitted to the atmosphere by respiration (mainly from soils), the share of C in dead material deposited over soils which is not fully degraded and mineralised back to CO₂ interacts with mineral soil particles (Rasmussen et al., 2018), and gradually builds up the soil organic matter (SOM), which is a pool with much higher residence times (centuries; Ciais et al., 2013). The degradation rate of litter depends on several factors, which in turn are mainly controlled by vegetation species, such as the litter quality (e.g., C, N, cellulose, and lignin contents) and quantity, the microbial community of decomposers, carbohydrate allocation, root biomass and surface soil temperature and moisture (Fernández-Alonso et al., 2018a). In particular, litter in beech forests is rich in lignin and cellulose content, and hence fungi dominate the microbial community because they can better use that substrate than bacteria (Schneider et al., 2010). However, once inside the soil, the interaction between SOM and small particles (clay; <2 µm) surfaces is complex and partially understood (Rasmussen et al., 2018). In general, more clay content gives more physical protection to SOM, promoting stabilisation and preventing C depletion (Hartley et al., 2021), but also the chemical composition of the SOM plays a role (Conant et al., 2011). Moreover, the van Bemmelen factor assumes that more than half of SOM is C (i.e. 58%; Howard & Howard, 1990). This soil organic carbon (SOC) is the largest terrestrial C pool, estimated at around 1700 Pg C, roughly thrice and twice as big as the vegetation and the atmospheric pool, respectively (Canadell et al., 2022). Likewise, global forests act as permanent net sinks of C, e.g. Pan et al. (2011) estimated a sink effect of temperate forests of 0.78 ± 0.09 Pg C yr⁻¹ for the period 2000-2007.

Climate change's impact on the land sink, and SOC, will depend on the future atmospheric CO₂ concentration scenarios. On the one hand, the land C sink is expected to increase with the atmospheric CO₂ concentration (fertilisation effect; Friedlingstein et al., 2021), which can lead to more SOC. However, the sequestration effect of land and soils can be reversed in the potential (and desired) scenario of decreased atmospheric CO₂ (Friedlingstein et al., 2021). On the other side, global warming enhances GPP (more C input) but also soil respiration (Rs) (Bond-Lamberty & Thomson, 2010), where there is no water limitation. Furthermore, the SOC decrease effect due to warming might be lower for fine-textured soils (clay protection effect; Hartley et al., 2021). Likewise, soil moisture content variability reduces the land C sink (Green et al., 2019). Nevertheless, the net effect of global warming on SOC is not clear yet (Davidson & Janssens, 2006; van Gestel et al., 2018), and the same applies to more frequent and intense droughts (Canadell et al., 2022). In addition, increased nitrogen deposition can increase C fixation in temperate forests ecosystems (see section 1.7), although it would be less important compared with CO₂ fertilisation and climate change effects (e.g., precipitation and temperature patterns changes; Zaehle & Dalmonech, 2011; Ciais et al., 2013; Fernández-Alonso et al., 2021).

Rs, performed by microorganisms and plant roots, releases vast amounts of CO₂ (~94 Pg C yr⁻¹), representing the largest C flux between land and the atmosphere after photosynthesis (Xu & Shang, 2016) and being almost ten times higher than fossil fuel emissions on an annual basis (Canadell et al., 2022). Part of the SOC is partially protected by complexation with clay-sized minerals or by aggregates occlusion (Hartley et al., 2021). However, microbes rapidly decompose the accessible part of SOC (Schmidt et al., 2011) and litter while plants roots respire photosynthates, and both processes combined conform to the soil respiration, and this is the main origin of the soil CO₂ efflux (Chemical oxidation and carbonate dissolution also exist; Lankreijer et al., 2003). Moreover, Rs represents between 60-80% of ecosystem respiration in temperate broad-leaved forests (Granier et al., 2003).

1.3. The nature of soils and their contribution to climate warming

The soil is a complex three-dimensional entity which consists of mineral and organic materials of varied sizes and chemical compositions. The finer the material and the more SOM content, the higher the ability to agglutinate and form porous corps, known as soil macro and micro aggregates (Six et al., 2004). The granular size composition, termed soil texture, together with the aggregation capacity, leads to the spatial arrangement of pores, termed soil structure, which defines essential soil functions like water holding or air capacity (Cosby et al., 1984; Moyano et al., 2013), both regulators of the biochemical reactions. Soil structure varies among different types of soils (e.g. agricultural vs forest soils) and soil horizons (e.g. superficial vs non-superficial), and so do the processes that depend on it. Furthermore, SOM association with clay-minerals to form microaggregates prevents its mineralization (Six et al., 2004).

1.4. Rs drivers

Soil temperature and moisture are primarily known to control Rs temporal variability (Davidson et al., 2006a; Bahn et al., 2009 & 2010), although other soil parameters that regulate microbial activity can also be important when spatial variability is considered. In particular, soil temperature explains most of the temporal variation but only when soil moisture is not too scarce or too abundant (Valentini et al., 2003; Bahn et al., 2010; Leitner et al., 2017; Fernández-Alonso et al., 2018b; Moyano et al., 2013).

1.4.1. Soil temperature

Microbial and plant roots activities are especially sensitive to temperature changes, usually showing a positive correlation (Janssens et al., 2003). Several equations have been used to describe the CO₂ relationship against temperature, as summarised in Janssens et al. (2003), who support the Arrhenius-type equation best describes the Rs-temperature relation. Other equations are linear, power, sigmoid, and exponential. The exponential equation Q₁₀ is often used, but the authors criticised it since it underestimated fluxes at low temperatures and overestimated them at high temperatures (Lloyd & Taylor, 1994). However, their fitting exercises showed that both Arrhenius and Q₁₀ relationships overestimated fluxes at high

temperatures from summer and, most importantly, coincided with lower soil moisture content. However, the performance of the models substantially improves when incorporating the moisture factor. Because of this, the authors highlighted the importance of manipulative experimentation (water exclusion and irrigation) under dry and warm conditions in the summer period to explore soil moisture scenarios that would be carried out later more extensively (e.g., Almagro et al., 2009; Bahn et al., 2010; Leitner et al., 2017; Fernández-Alonso et al., 2018 & 2021).

1.4.2. Soil moisture

Soil moisture is the main factor explaining soil CO₂ efflux when it is scarce. This has been described for several forests ecosystems (Epron et al., 1999; Curiel Yuste et al., 2003; Almagro et al., 2008; Luo et al., 2012; Leitner et al., 2017; Fernández-Alonso et al., 2018). Moreover, an exacerbated reaction has been described when the soil water content is recovered after a dry period (Almagro et al., 2008; Birch, 1958; Curiel Yuste et al., 2003 & 2007; Fierer & Schimel, 2002; Jarvis et al., 2007; Miller et al., 2005; Orchard & Cook, 1983; Xiang et al., 2008), sometimes leading to higher soil CO₂ release compared with continued moisture conditions. This, combined with the projection of an increase in frequency and intensity of DRW events, would positively feedback climate change.

1.5. Effect of DRW cycles on soil and Rs

1.5.1. Drought

A prolonged period without water input dries the soil below the field capacity, modifying several physical and biological processes.

While water film gets disconnected, it reduces the nutrient transport (Skopp et al., 1990), isolating and concentrating the nutrients in smaller pores. Structure stability declines in coarser textures while increases in finer soils. Besides, the extent of shrinking provoked by drying depends on the type and amount of clay. Also, Bimüller et al. (2014) reported a hindered soil N processing (i. e., mineralisation) and posterior stabilisation in organo-mineral fractions during drought, with the potential risk of loss of labile N in extreme rainfall events.

Extremely dry conditions trigger soil microbiota's specific reactions (Borken & Matzner, 2009; Barnard et al., 2020). CO₂ fluxes are highly reduced during dry conditions (Orchard & Cook, 1983; Schimel et al., 1999; Howard & Howard, 1993;). The decrease in microbial activity might explain this, and even death, caused by limited water availability (De Nobili et al., 2006). One of the strategies to avoid death by desiccation is concentrating osmolytes (Harris, 1981). Sometimes, forming spores and undergoing dormancy is a better survival strategy. Exudated enzymes maintain active mineralising C and N during drought (Manzoni et al., 2014). The longer the drought period, the greater the impact on the microbial biomass (Schimel et al., 1999), and any resistance strategy represents a cost in energy and nutrients (Schimel et al.,

2007). Lastly, drought promotes hydrophobicity, but Manzoni et al. (2014) considered it and the aggregate stability less important than osmotic regulation and nutrient diffusion for their impact on microbial activity.

1.5.2. Rewetting and “Birch effect”

Rewetting dry soils usually triggers a disproportionated burst of CO₂, a phenomenon known as the “Birch effect” (Jarvis et al., 2007), that would be decoupled from microbial growth (Göransson et al., 2013; Iovieno and Bååth, 2008). Any considerable amount of water addition changes the water potential and has more complex effects than just refilling the soil. The microbial activity recovers sharply after a substantial rewetting, often more than expected in the hypothetical situation of continuous moist soil (Birch, 1958a and b; Fierer & Schimel, 2002; Iovieno and Bååth, 2008; Orchard & Cook, 1983). Recovery after rewetting can take minutes (Borken et al., 2003), but it can also be delayed by dormancy, from 1 to 72 hours (Placella et al., 2012). The delay depends on the drought length (Barnard et al., 2015; Göransson et al., 2013) and harshness (Meisner et al., 2017), the adaptation of microbes to DRW stress (Meisner et al., 2015), and the history of past DRW events with the possible adaptation of microbiota after repeated cycles (Fierer & Schimel, 2002; Leizeaga et al., 2021 & 2022).

Researchers proposed different mechanisms to explain the “Birch effect” (Barnard et al., 2020; Borken & Matzner, 2009; Evans et al., 2006; Fierer & Schimel, 2003; Manzoni et al., 2014). Microbes need to release the osmolytes accumulated during dry conditions (e.g. proline) the better as possible to avoid plasmolysis, and these components would be quickly mineralised. However, because the turnover time is short (Warren, 2019), it is hard to demonstrate that released osmolytes actually provide a source of C. Hence, contradictory results exist (Boot et al., 2013; Warren, 2014; Williams & Xia, 2009). Microbial death, either by plasmolysis (Kief et al., 1987) or infection by phages and viruses (Williamson et al., 2017), also might contribute as C source. Water film disconnection due to dryness promotes microbial diversity (Carson et al., 2010), which provides higher adaptability and possibly faster recovery when the porosity film connection returns. The persistence of activated exoenzymes is another proposed explanation (Lawrence et al., 2009). However, Homyak et al. (2018) did not find evidence of persistent exoenzyme activity in dry soils. Clay complexes can release organic matter by desorption (Blankinship & Schimel, 2018). The disruption of aggregates releases occluded organic matter (Cosentino et al., 2006), which would also be an extra C substrate upon rewetting (Navarro-García et al., 2011). When DRW stress releases previously unavailable labile organic compounds (Schimel et al., 2011), the pulse of CO₂ is higher than in continually moist soil (Borken & Matzner, 2009). Nevertheless, the exact mechanistic processes of the DRW cycles are still under debate, and hence also, its effects and interactions with climate change. Furthermore, manipulative field experiments that simulate DRW cycles in temperate forests are lacking.

1.6. Other drivers of Rs - Spatial variability

Rs also shows large spatial variability even within short distances, especially under forest (Bahn et al., 2008; Curiel Yuste et al., 2004; Epron et al., 2004b; Kang et al., 2003; Law et al., 2001; Sørensen & Buchmann, 2005; Vincent et al., 2006), which can be higher than temporal variability (at least than diurnal variation; Janssens et al., 1998; Law et al., 2001; Rayment & Jarvis, 2000), and can be explained by several soil parameters. As discussed above, soil temperature and moisture are strong predictors of temporal variations, but the explanation of spatial heterogeneity in fluxes might require including other soil parameters. Among those factors, Vincent et al. (2006) have mentioned vegetation (Law et al., 2001) and its roots properties (Janssens et al., 1998), organic matter quantity and quality (Epron et al., 2004b; Rayment & Jarvis, 2000), microbial biomass (Lee & Jose, 2003), and soil texture and the related porosity (Dilustro et al., 2005; Moyano et al., 2012). Furthermore, soil chemistry has also been found to explain Rs in temperate forests, and examples are Janssens et al. (2003) and Borken et al. (2002), both reporting negative correlation with C:N ratios, or Vincent et al. (2006) showing correlations with topsoil N content. In addition, Sørensen & Buchmann (2005) found a positive correlation of Rs with several soil chemistry parameters (C:N and concentrations C_{tot} , N_{tot} , P, DOC, among others) and soil structure and roots characterisation parameters. Likewise, Kang et al. (2003) reported that CO₂ efflux variability within short distances could be better explained by soil moisture than temperature. However, more attention has been focused on Rs's temporal rather than spatial drivers.

1.7. Atmospheric N deposition effect on soil CO₂ efflux

Twentieth-century industrial fixation of N₂, inert and abundant (80%) in the atmosphere, into reactive N (Nr) forms (ammonia, nitrates, amino acids, proteins, among others) for posterior release in ecosystems resulted in a “cascade” of intended and unintended consequences for the C cycle still unclear (Arias et al., 2021; Sutton et al., 2011). Satisfying N-demand for food production is a positive effect. On the contrary, a harmful example is that higher emissions of NO_x and NH_x derived from human high-temperature combustion processes and gaseous loss from N-enriched ecosystems have increased the atmospheric Nr deposition (Sutton et al., 2011). In Europe, although deposition in oxidised form (NO_x) has peaked in the 1980s and then halved by 2017, it is still 3-4 times higher than 1900 levels, while the value for the reduced form (NH_x) doubles the 1900 levels and it is not expected to decrease by 2050 (Engardt et al., 2017). Among several effects, the increased Nr over temperate forests has been generally described as reducing Rs and increasing C sequestration (Dirnböck et al., 2017; Janssens et al., 2010; Pregitzer et al., 2008; Zhou et al., 2014; Zhong et al., 2016), with the magnitude moderated by soil conditions, such as previous Nr availability (Dirnböck et al., 2017), or MBC, SOC, bulk density, and pH (Zhong et al., 2016). While dry conditions can also reduce Rs, hampering microbial activity by water limitation (Fernández-Alonso et al., 2021), the interaction between chronic atmospheric Nr deposition under more intense and frequent DRW cycles has not been adequately assessed yet.

1.8. Objectives

This study aims to assess the effect of drying-rewetting stress on the soil CO₂ fluxes and to uncover the controlling soil parameters in one mature European beech (*Fagus sylvatica* L.) forest in Eastern Austria during one drying-rewetting stress event, under different nitrogen deposition scenarios.

Specific objectives

- a) We will assess whether potential future intensified summer DRW stress in temperate forests enhances or hampers soil CO₂ efflux.
- b) We will describe the effect of increased atmospheric N deposition on the soil CO₂ efflux under DRW stress.
- c) Exploring the explanatory power of soil parameters will improve mechanistic understanding of any response of CO₂ efflux to the changing environmental conditions.
- d) We will describe the spatial variability of soil CO₂ efflux, identifying possible emission “hot spots”.
- e) We will try to identify if the “Birch effect” occurs under the specific soil moisture of a temperate forest during a summer drought.

1.9. Hypothesis

- i) Since rewetting the dry soil expectedly reactivates the microbial activity, soil CO₂ efflux and MBC would rise after irrigation.
- ii) If, upon rewetting, the soil CO₂ efflux gets decoupled from soil moisture, meaning there is an exaggerated reaction considering the soil moisture level, it would be attributed to the Birch effect occurrence.
- iii) Soil temperature and moisture are strong predictors of Rs, but the latter may be more important in dry conditions. Consequently, the soil moisture would explain the variability in CO₂ efflux better than soil temperature and positively correlate with the CO₂ efflux in the DRW treatment.
- iv) Forests soils show high variability of soil biochemical and physical parameters. If this is the case, CO₂ efflux would also show high spatial variability and correlate with any of the biochemical and physical soil parameters monitored.
- v) We expect that enrichment of forest soils with Nr will increase the concentration of mineral forms of N, either NO₃⁻ or NH₄⁺, reduce microbial activity (MBC), and suppress soil CO₂ efflux.

2. Materials & Methods

2.1. Experimental Site

We recovered field data from a “Beech” (*Fagus sylvatica* L.) forest stand in Klausen-Leopoldsdorf. This long-term ecosystem research (LTER) site is managed by Federal Research Centre for Forests, Natural Hazards, and Landscape (BFW), and it includes varied research equipment, such as rain-out shelters and an irrigation system automated chambers for soil-atmosphere gas exchange measurements, and climatic sensors.

It is situated in a managed homogenous beech stand, 80 years old, and located 20 km west of Vienna (48°07' N 16°03' E). Since 2006, it has been part of the LTER Austria network. It is also part of the European Level II Forest Monitoring System in the International Co-operative Programme on Assessment and Monitoring of Air Pollution Effects on Forests (ICP Forests). According to Kitzler et al. (2006), the soils are Dystric Cambisols developed over sandstone (WRB), with a loam-loamy clay texture, and the natural N deposition rate is around 13 kg N ha⁻¹ y⁻¹. At the plot's site, the elevation is 500 m a. s. l., the slope is exposed to NNE, and the mean annual temperature and precipitation are 8°C and 728 mm. Bulk density is 0.827 g cm⁻³. Finally, Table 2-1 summarises the essential features of the site.

Table 2-1. Main features of site Klausen-Leopoldsdorf. Data based on Kitzler et al. (2006).

Klausen-Leopoldsdorf	
GPS Coordinates	48°07' N 16°03' E
Elevation (m a.s.l)	500
Annual mean temperature (°C)	8
Annual mean precipitation (mm)	728
Mean pp on vegetation growing period (mm, 1970-2000)	446
Soil classification (WRB)	Dystric Cambisols over Sandstone
Nitrogen deposition rate (kg N ha ⁻¹ y ⁻¹)	13
Hill exposition and inclination	NNE
Soil texture	Loam-Loamy clay

2.2. Timespan and experimental design

The year 2021 was the first of the 3-years-EXAFOR project, which consisted of field monitoring of soil GHGs fluxes and several other associated parameters under a rainfall exclusion experimentation combined with wet N fertilization at two research forest sites, “Klausen-Leopoldsdorf” and “Rosalia”. From April till November, covering the vegetation growing period, 3 complete severe DRW cycles were simulated, each of them consisting on 8 weeks of drought without water inputs interrupted by an irrigation event of 150 mm. Only the time distribution of water inputs and not the natural average total amount was manipulated. The 3 irrigation events approximately sum up to the average precipitation recorded for the vegetation growing season for the years 1970 to 2000 (Table 2-1; Kitzler et al., 2006). At Klausen-Leopoldsdorf, the rainfall exclusion started on May 4th and ended on November 16th. However, for the current thesis work, only the data for the 28-days-period between August 19th

and September 15th. This includes the last 6 days of the second drought, the second irrigation event of August 25th, and 22 days more after the rewetting until September 15th (Figure 2-1).

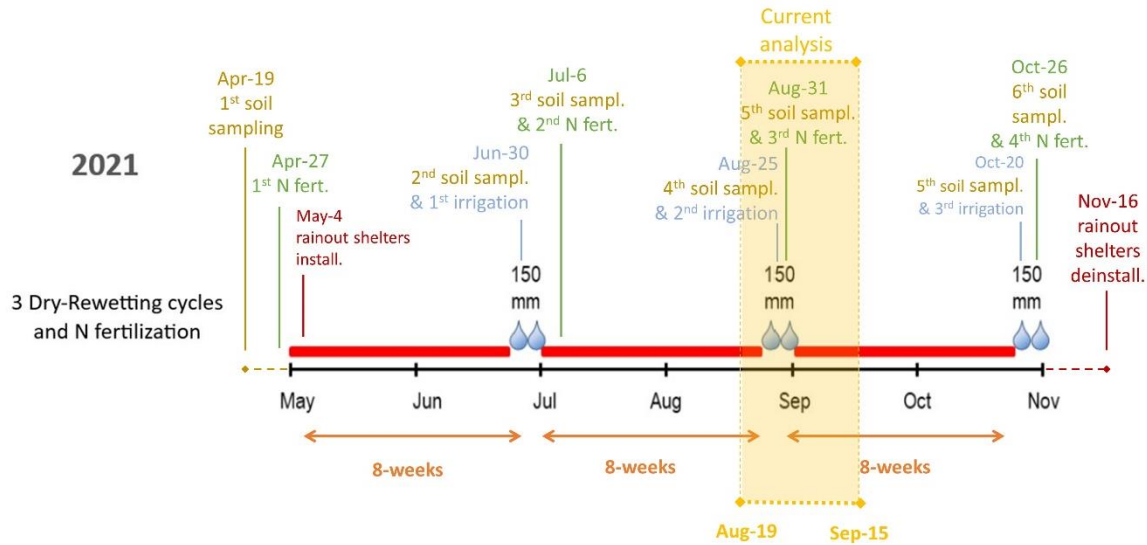


Figure 2-1. Timeplan of the year 2021 at Klausen-Leopoldsdorf research site. Adapted from Institute of Soil Research, BOKU.

The experimental setup included 16 square plots (3x3 m) allocated along a slope (Figure 2-2). The experimental design consisted of a factorial approach, combining two levels of rainfall manipulation (Control and severe DRW stress) with two other N addition scenarios (natural and +50 kg N ha⁻¹ a⁻¹). We replicated randomly assigned four plots for each of the four resulting combination treatments (“C”, “C + N”, “S”, and “S + N”). The plots distribution resulted in: Plots number 1, 5, 10, and 13 assigned to “Control” (C) treatment; plots 2, 6, 9, and 14 to “Control with N addition” (C+N); plots 3, 7, 11, and 15 to “Severe DRW stress” (S); plots 4, 8, 12, and 16 to “Severe DRW stress with N addition” (S+N). Consequently, half of the plots were sheltered by May 4th.

2.3. Rainfall manipulation

Rain-out shelters in combination with an irrigation system were used to simulate severe DRW stress in half of the plots (Figure 2-2). The shelters were installed from May 4th till November 16th (Figure 2-1). Appendix A includes photos of the shelters with the irrigation system. At every irrigation event, the volume of water each plot required was:

$$(0.150\text{m} \times 3\text{m} \times 3\text{m}) \text{ plot}^{-1} = 1.35 \text{ m}^3 \text{ plot}^{-1} \times 1000 \text{ L m}^{-3} = 1350 \text{ L plot}^{-1}$$

Which throws a total of:

$$1350 \text{ L plot}^{-1} \times 16 \text{ plot} = 21600 \text{ L}$$

A plastic container bag of at least 22 m³ (22000 L) was placed at a point of the slope of a higher level of all the setup and filled using bombs. Furthermore, the water had to be decalcified to avoid unwanted C and Ca²⁺ inputs to the soil, which could interfere with the treatments. For that purpose, a descender system was implemented. The water flowed by the difference in

gravitational potential through a flexible line which ended at a 1st transfer 1m³-container. From the 1st container, the water was pumped to the descender and then transferred to a second 1m³ container from which the water was pumped and directed through flexible lines to the irrigation systems mounted at each sheltered plot. Below each shelter, a distribution rigid pipeline system was installed, with a valve and a manometer at the inlet point and water aspersion at several points in the lines. The water pressure was monitored and maintained at a level which allowed homogeneous aspersion over the soil surface.

Klausenleopoldsdorf Site – Overview Status 2021



Institute of Soil Research

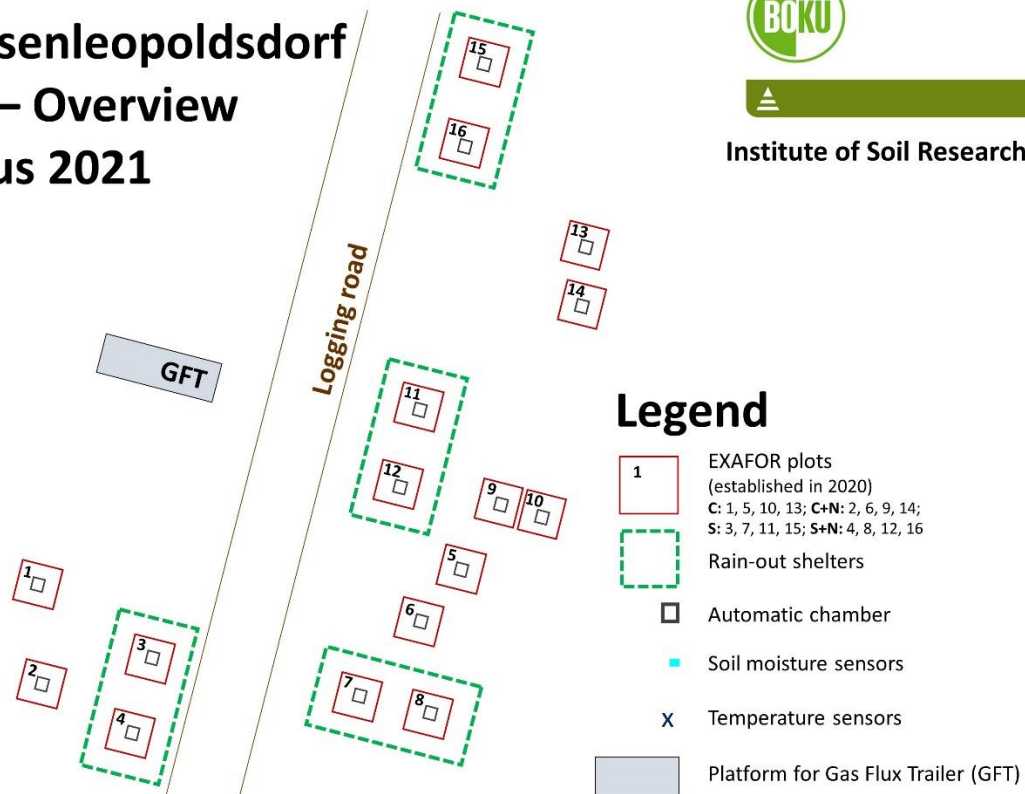


Figure 2-2. Experimental setup at Klausen-Leopoldsdorf research site. Each of the 16 squared plots (3 m x 3 m; red squares), was equipped with an automated chamber (black squares) connected to a “Gas Flux Trailer” (grey rectangle). Half of the plots were covered with rain-out shelters (green dashed rectangles) since May 4th, 2021. The plots were randomly assigned and equally distributed to each of the four treatments resulted. Plots number 1, 5, 10, and 13 were assigned to Control (C) treatment; 2, 6, 9, and 14 to Control with N addition (C+N); 3, 7, 11, and 15 to Severe DRW stress (S); 4, 8, 12, and 16 to Severe DRW stress with N addition (S+N). Source: Institute of Soil Research, BOKU.

2.4. N fertilization

A scenario of increased atmospheric N deposition was simulated by N fertilization in half of the plots at a rate of 50 kg N ha⁻¹ y⁻¹. The N addition was subdivided into 4 N fertilization events (Figure 2-1), consisting of 12 kg N ha⁻¹. The amount added at each event for each plot was:

$$12 \text{ kg N ha}^{-1} (10000 \text{ m}^2 \text{ ha}^{-1})^{-1} = 0.0012 \text{ kg N m}^{-2}$$

Multiplied by the area of the plots:

$$0.0012 \text{ kg N m}^{-2} (3 \text{ m} \times 3 \text{ m}) \text{ plot}^{-1} = 0.0108 \text{ kg N plot}^{-1} = 10.8 \text{ g N plot}^{-1}$$

A 20 L solution of 0.54 g N L⁻¹ concentration was prepared with ammonium nitrate (NH₄NO₃) granular fertilizer and evenly applied in two rounds of 10 L each, with the use of 12 L-capacity backpack sprayers (Appendix A). The addition of 20 L to the plots represented a water input event of:

$$20 \text{ L} (1000 \text{ L m}^{-3})^{-1} (9 \text{ m}^2)^{-1} = 0.002 \text{ m} = 2 \text{ mm}$$

To compensate for the water applied in the fertilized plots, the unfertilized plots also received an equivalent application of water.

2.5. Soil CO₂ flux estimation

As shown in Figure 2-2, one automated chamber (non-steady-state non-flow-through; Livingston and Hutchinson, 1995) was placed at the centre of each plot. The chambers used were “black” type because their structure was made of stainless steel, and this material prevents sunlight from entering, so the light-dependent part of photosynthesis is stopped while the chamber is closed. The chambers were installed over stainless-steel frames inserted 5 cm in the mineral soil, and their dimensions were 0.5 m x 0.5 m (horizontal sides) x 0.15 m (height). Each chamber also contained one fan, which ensures homogeneous air mixing. The 16 chambers were connected to the “GasFluxTrailer”, which controlled the opening and closing and the gas sampling timing and contained a cavity-ring-down spectroscope which measures the CO₂ air concentration (Picarro G2301, Santa Clara, USA), among other required equipment (computer, data logger, another spectroscope for measuring N₂O, etc.). The temporal resolution of gas exchange measurements was three hours, while the chamber enclosure time was 10 minutes.

2.5.1. Flux calculation

Once gas concentration inside the chamber is estimated over the 10 minutes enclosure time, the several measured values for the gas concentration inside the chamber combined with the exact measurement time are used to estimate the temporal rate of change of the concentration. This change can be linear or non-linear. Non-linear relations are attributed to artefacts in the chamber technique, but several corrections have been suggested (Parkin & Venterea, 2010). In the case of a constant rate of change, a linear regression can be used to calculate the slope, which would be expressed as, e. g. [μL CO₂-C L⁻¹ min⁻¹] or [ppmv min⁻¹]. Due to the relative short enclosure time and large chamber size, the change of rate was calculated with linear regression.

Following Butterbach-Bahl et al. (2011), the steps for the flux of CO₂ [μg C m⁻² h⁻¹] calculation are:

$$\text{Flux of CO}_2 (\mu\text{g C m}^{-2} \text{ h}^{-1}) = \frac{\text{chamber volume (m}^3\text{)} \text{ mol weight of C (g mol}^{-1}\text{)} \text{ regression slope (ppmv min}^{-1}\text{)} 60 \text{ min } 10^6 \mu\text{g}}{\text{chamber area (m}^2\text{)} \text{ mol volume of gas (m}^3 \text{ mol}^{-1}\text{)} \text{ hour g}}$$

But, the molar volume of gas needs to be corrected according to the ideal gas law:

$$\text{mol volume of gas (m}^3 \text{ mol}^{-1}\text{)} = 0.02241 \frac{(273.15 + \text{Temp (}^\circ\text{C)})}{273.15} \frac{\text{atmospheric pressure at measurement height (Pa)}}{\text{atmospheric pressure at sea level (Pa)}}$$

Atmospheric pressure can be estimated with height above sea level with a barometric formula (Butterbach-Bahl et al., 2011), and headspace air temperature has to be measured at the enclosure time.

The unit presented in the current study is mg CO₂-C m⁻² d⁻¹, hence two more transformations are needed:

$$\text{mg CO}_2\text{-C m}^{-2} \text{ d}^{-1} = \mu\text{g CO}_2\text{-C m}^{-2} \text{ d}^{-1} (10^3 \mu\text{g mg}^{-1})^{-1}$$

Finally, the measurement was done every 3 hours per chamber, but the 8 intraday values were pooled into one daily average.

2.6. Soil sampling

On each sampling date (Figure 2-1), a regular metal mesh of 1.5 m long and 1 m wide, and rectangular cells (5 cm x 5 cm), with columns and rows rotulated, was placed inside the plots on the right side of the chamber. Five cells defining the soil auger (1 cm diameter) entrance point were randomly selected at each sampling date and discarded for the remaining period. Hence, the auger was introduced 5 times up to 20 cm inside mineral soil after manual litter removal, without repeating the cell in the entire 1-year-campaign. The material recovered inside each auger was divided by depth, from 0-10 and 10-20 cm. The five soil samples from the same depth were pooled, sieved (2 mm) and enclosed in bags. At this point, we weighted each bag (2 per plot) to ensure we contained 20 g of soil at a minimum, and if needed, one extra augering at a random point was performed. In the end, the two bags were adequately closed and refrigerated for posterior transport to the laboratory. This procedure was repeated in the 16 plots.

The current thesis includes the 4th (Aug 25th) and 5th (Aug 31st) soil sampling campaigns (Figure 2-1).

2.6.1. Laboratory analysis

At the lab, we analysed the soil samples to obtain soil texture and moisture and the concentrations of microbial biomass carbon (MBC) and nitrogen (MBN), dissolved organic carbon (DOC), total dissolved nitrogen (TDN), ammonium (NH₄⁺), and nitrate (NO₃⁻).

By the Chloroform-Extraction-Method (Schinner and Sonnentner, 1996), we obtained the microbial biomass carbon (MBC) and nitrogen (MBN).

We measured in a carbon and nitrogen total analyser (TOC-L, Shimadzu, Kyoto, Japan) the total dissolved carbon (TDC) and nitrogen (TDN) after extracting with 0.5 M K₂SO₄ solution, 30-min shaking, and N-free filtering. We read NH₄⁺ and NO₃⁻ following Jones and Willett (2006), which also allowed us to estimate dissolved organic nitrogen (DON) by subtraction.

The non-fumigated extract allowed us to estimate the dissolved organic carbon (DOC) and nitrogen (TDN).

Soil texture was obtained by sedimentation; We weighted the samples before and after oven drying (105 °C) to estimate soil moisture content.

2.7. Meteorological data

The Gas Flux trailer has installed a tipping bucket for estimating precipitation and an air temperature sensor located 2 m above ground. Both types of equipment have a temporal resolution of 1 minute.

In addition, BFW provided data for the following parameters: Soil temperature at 5 cm depth, measured with thermocouples; Soil water content estimated by water content reflectometer. The time resolution of this data was 30 min. However, sensors were installed only in 4 plots, 2 sheltered and 2 controls.

Figure 2-3 and Figure 2-4 show the daily average soil moisture, precipitation, and irrigation events, while Figure 2-5 resumes the temperature data.

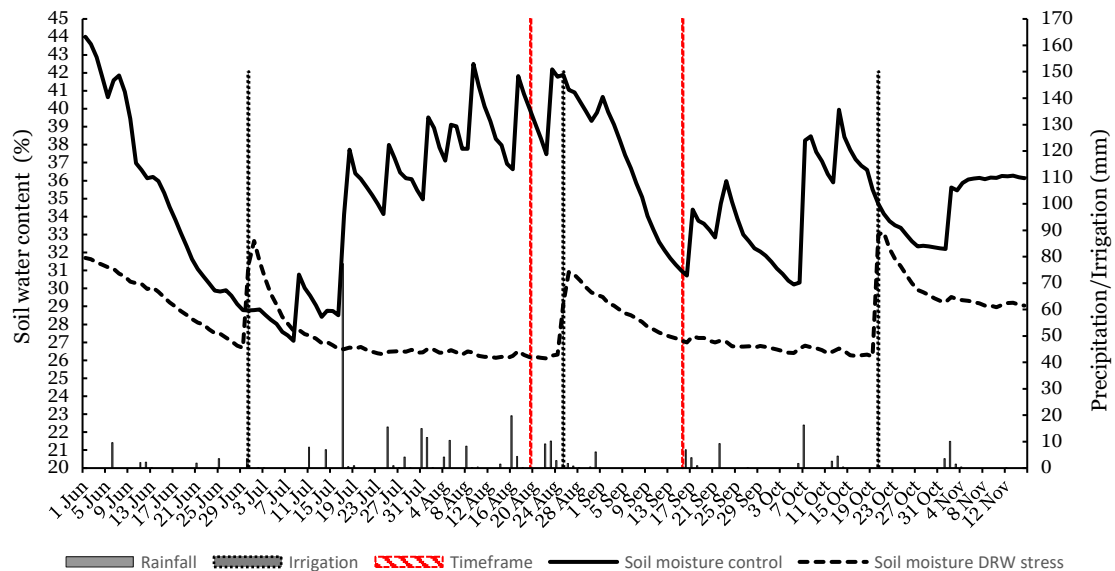


Figure 2-3. Evolution of daily averages for soil water content (%) (left axis) and water inputs (mm) (right axis) between June 1st and November 12th. The soil water content is differentiated by DRW (dashed line) and control (continued line) plots. The water inputs are differentiated by precipitation (black columns) and irrigation (dashed columns). The data analysed in the current thesis falls between the red lines.

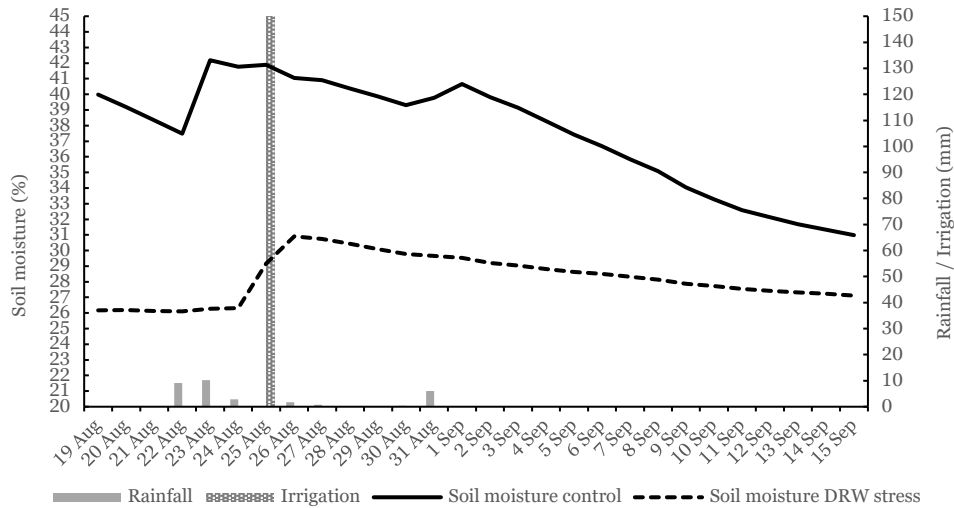


Figure 2-4. Evolution of daily averages for soil water content (%) (left axis) and water inputs (mm) (right axis) between August 19th and September 15th. The soil water content is differentiated by DRW (dashed line) and control (continued line) plots. The water inputs are differentiated by precipitation (solid columns) and irrigation (dotted columns).

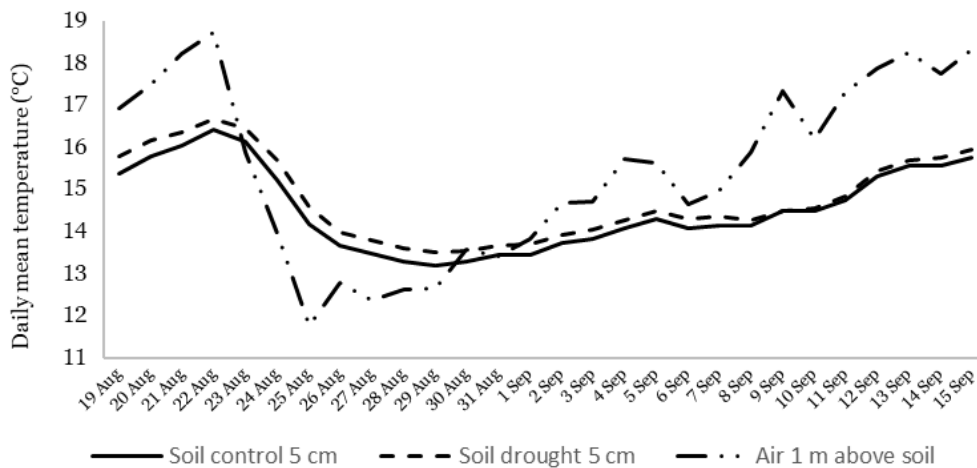


Figure 2-5. Evolution of daily averaged temperature between August 19th and September 15th, at 5 cm soil depth in control (solid line) and DRW stress (dashed line), and air temperature at 2 m above soil (dashed and pointed line). “drought” is referring to “DRW stress”.

2.8. Data handling and statistical analysis

Software

For creating data frames, tables and figures, Microsoft Excel and RStudio were used (R Core Team, 2021; RStudio Team, 2021), but only the latter was used for the statistical analysis. Contour plots were performed with Surfer[®] (Golden Software, LLC).

Transformations

The most simple transformations were pooling and averaging the data to obtain daily means. This was the case for data with time resolution lower than intraday level, like CO₂ fluxes, soil

moisture and temperature, air temperature, and precipitation. Since the soil sensors were installed 2 at sheltered and 2 at non-sheltered plots, it was necessary to pool this data into DRW stress and control treatments for posterior correlation and linear regression analysis with CO₂ fluxes.

However, some data was also log-transformed (\log_{10}) to achieve normality and reduce the effect of outliers, and this was the case for CO₂ efflux, and for many ancillary physicochemical soil parameters, before mean comparisons, correlation, or regression analysis.

For multiple mean comparisons, i. e. when the 4 combination treatments (C, C+N, S, S+N) were compared, the first option was ANOVA, followed by Tukey's post hoc analysis. However, if the normality assumption could not be fulfilled, the Kruskal-Wallis test was used, followed by Wilcoxon's sum rank test (with Bonferroni correction) as post hoc analysis.

Validation

The validation approach of models was based on Zuur et al. (2007), and heteroscedasticity was tested by plotting the models' residuals against fitted values. A QQ-plot of the residuals was used for normality and residuals against every explanatory variable for independence.

3. Results

3.1. Spatial variability

The boxplots shown in Figure 3-1 were constructed using combining flux data of two days (August 24th and 31st). The resultant pattern of CO₂ efflux across the plots is typical for the period analyzed and gives an idea of the spatial variability of the setup. A Kruskal-Wallis test was performed to explore differences by treatment using the two days fluxes, and although it was significant ($P<0.05$), Wilcoxon's sum rank test with Bonferroni's correction gave no difference between treatments. The mean and SD value for the 2 days was 228.0 ± 155.9 CO₂-C m⁻² h⁻¹. If days consecutive days around the two dates are averaged, e. g. the sampling date and the 4 previous days, from August 20th to 24th (before rewetting) and from August 27th to 31st (after rewetting), the average flux and SD now is 229.7 ± 152.4 CO₂-C m⁻² h⁻¹, almost the same result.

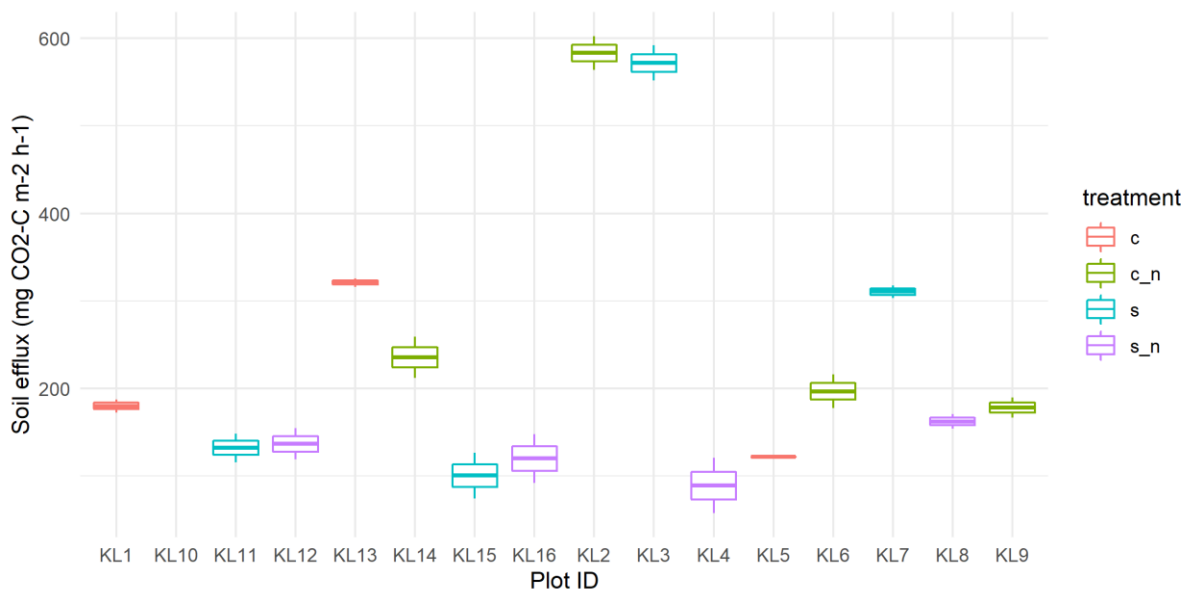


Figure 3-1. Boxplots of daily average soil efflux (CO₂-C m⁻² h⁻¹) from each plot, and coloured by the 4 treatments combination. Boxplots contain only data from August 24th and 31st.

For further assessing the contribution of the spatial variability in the soil CO₂ efflux, the ancillary soil data were grouped and compared by sampling date, before (August 24th) and after (August 31st), and by treatments (either by pp exclusion*N addition or only by pp exclusion). The significant results showed that the total N at 20 cm was higher ($P<0.05$) and the C:N ratio at 20 cm lower ($P<0.05$) for the DRW stress compared with control. Also, the lower C:N ratio at 20 cm value was for the “S” treatment. Also, the microbial biomass N was lower ($P<0.05$) for the DRW stress plots, but only on the first sampling date. The contrary happened with ammonium, being higher for the DRW plots but only at the second sampling date ($P<0.05$). The comparisons between the 4 treatments combined (C, C+N, S, and S+N) resulted in all the ANOVA and Kruskal-Wallis tests not being significant. The summarized data and p values are shown in Table 0-1(Appendix A).

On the other hand, the soil efflux data was correlated and linear regressions were fitted with the soil parameters without any differentiation by date or treatment and the results are showed in Table 2-1Table 3-1. Significant Spearman's correlations were found for MBC at 10 cm ($\rho = 0.55$; $P < 0.01$) and at 20 cm depth ($\rho = 0.4$; $P < 0.05$), for MBN at 10 cm ($\rho = 0.5$; $P < 0.01$) and at 20 cm ($\rho = 0.43$; $P < 0.05$), and for DOC at 10 cm ($\rho = 0.39$; $P < 0.05$). Furthermore, the significant linear regression were fitted for Total C at 20 cm ($R^2 = 0.14$; $P < 0.05$; after log transforming), for MBC at 10 cm ($R^2 = 0.16$; $P < 0.05$), for MBN at 10 cm ($R^2 = 0.16$; $P < 0.05$), and for DOC at 10 cm ($R^2 = 0.14$; $P < 0.05$). Lastly, a multiple linear regression which included the 4 variables of the simple linear regressions as explanatory variables, was almost significant ($R^2 = 0.19$; $P < 0.065$; not included in the table).

Table 3-1. Soil parameters mean and SD. Rho Spearman's correlation and R^2 linear regression were calculated, relating the variables with the log-transformed (\log_{10}) CO₂ efflux. Significance: * = $P < 0.05$; ** = $P < 0.01$. Transformation of the explanatory variables is indicated. Abbreviations: dw = soil dry weight; SD = standard deviation.

Parameter	Unit	Mean	SD	rho	R ²	Transformation?
<i>Sand (10 cm)</i>	%	13.4	3.43	0.30		
<i>Sand (20 cm)</i>	%	11	1.9	0.14		
<i>Silt (10 cm)</i>	%	55.2	4.36	<0.01		
<i>Silt (20 cm)</i>	%	54.4	3	<0.01		
<i>Clay (10 cm)</i>	%	31.42	4	-0.24		
<i>Clay (20 cm)</i>	%	34.8	2.9	-0.11		
<i>Total N (10cm)</i>	w/w%	0.19	0.06	0.09		
<i>Total N (20cm)</i>	w/w%	0.13	0.05	0.18		
<i>Total C (10cm)</i>	w/w%	3.6	0.8	0.04		
<i>Total C (20cm)</i>	w/w%	2.4	0.8	0.09	0.14*	Log
<i>C:N (10cm)</i>	-	19.4	3.2	-0.27		
<i>C:N (20cm)</i>	-	20.1	3.5	0.10		
<i>Microbial biomass C (10 cm)</i>	C mg (kg dw) ⁻¹	144.6	43.51	0.55**	0.16*	
<i>Microbial biomass C (20 cm)</i>	C mg (kg dw) ⁻¹	144.6	69.2	0.4*		
<i>Microbial biomass N (10 cm)</i>	N mg (kg dw) ⁻¹	19.2	6.2	0.5**	0.16*	
<i>Microbial biomass N (20 cm)</i>	N mg (kg dw) ⁻¹	20.2	9.5	0.43*		
<i>Dissolved organic C (10 cm)</i>	C mg (kg dw) ⁻¹	207.7	35.4	0.39*	0.14*	
<i>Dissolved organic C (20 cm)</i>	C mg (kg dw) ⁻¹	186	39	0.30		
<i>Dissolved organic N (10 cm)</i>	N mg (kg dw) ⁻¹	25.9	5.6	0.29		
<i>Dissolved organic N (20 cm)</i>	N mg (kg dw) ⁻¹	22.8	4.5	0.41		
<i>NO₃⁻ (10 cm)</i>	µg NO ₃ ⁻ -N (g dw) ⁻¹	2.6	2.2	0.20		
<i>NO₃⁻ (20 cm)</i>	µg NO ₃ ⁻ -N (g dw) ⁻¹	1.45	0.8	0.20		
<i>NH₄⁺ (10 cm)</i>	µg NH ₄ ⁺ -N (g dw) ⁻¹	7.3	2.9	-0.04		
<i>NH₄⁺ (20 cm)</i>	µg NH ₄ ⁺ -N (g dw) ⁻¹	5.6	1.8	0.05		

3.2. Temporal variability

Figure 3-2 and Figure 3-1 show similar patterns, but the new one includes all the average daily CO₂ efflux observations for each chamber, which is expressed as more vertical dispersion of the points and boxplots. Also, the overall mean and SD values were 222.5 ± 139.7 CO₂-C m⁻² h⁻¹, showing the same or lower level of variation than the spatial analysis and similar mean value. However, for this case, the Kruskal-Wallis test resulted in significant ($P < 0.0001$) for all the periods analysed, and the Welch's test when comparing before and after rewetting means ($P < 0.01$ at least). Wilcoxon's rank sum test showed treatments C+N and S with the highest mean flux, C with intermediate values, and S+N with the lowest (Table 3-2) for the three periods analyzed.

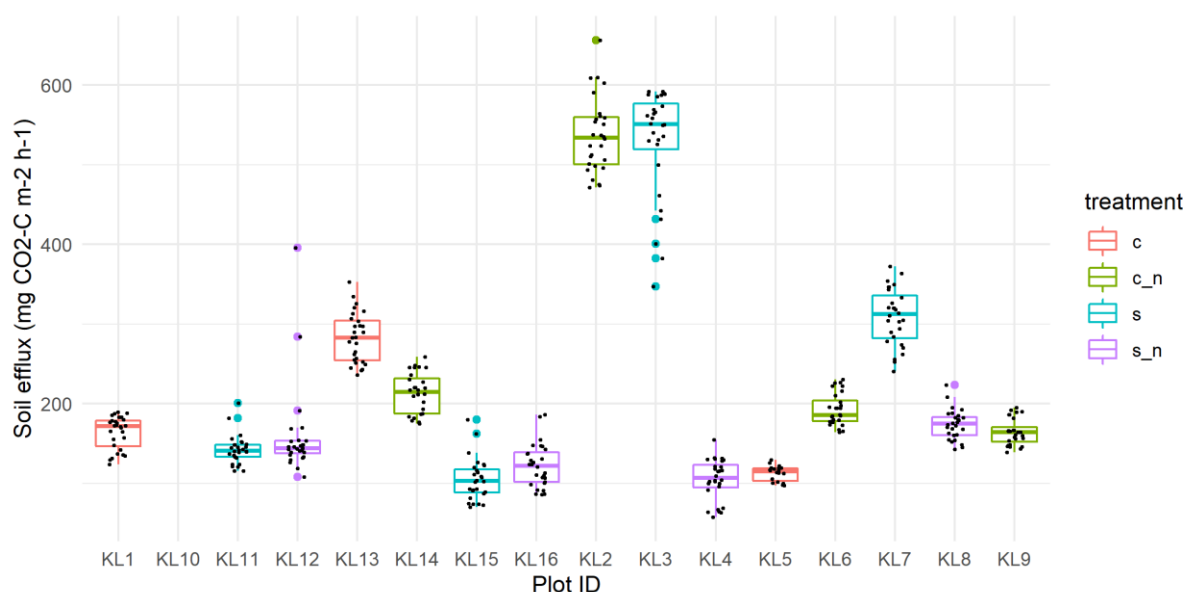


Figure 3-2. Boxplots of daily average soil efflux (CO₂-C m⁻² h⁻¹) from each plot and coloured by the 4 treatments combination. The data for the full period (28 days) is included. The points in black are all the flux observations (28).

Table 3-2. Mean \pm SE (min-max) of averaged CO₂ fluxes along days of the four treatments when including N addition levels. Three periods were considered, complete (28 days from August 19th to September 15th), before rewetting (6 days from August 19th to 24th), and after rewetting (22 days from August 25th to September 15th). The p-values inform the Kruskal-Wallis test. The letters above the means indicate ranks from Wilcoxon's sum rank test with Bonferroni correction.

	mg CO ₂ -C m ⁻² h ⁻¹				p-value
	C	C + N	S	S + N	
complete (28 d)	^b 200.14 \pm 7.20 (154.95-287.41)	^a 276.69 \pm 4.67 (241.99-329.3)	^a 271.25 \pm 4.41 (232.29-306.08)	^c 140.37 \pm 5.23 (103.22-226.10)	<0.0001
before rewett.(6 d)	^a 256.52 \pm 8.82 (211.29-287.41)	^a 310.42 \pm 5.62 (289.06-329.3)	^a 263.37 \pm 7.84 (244.33-306.08)	^b 127.98 \pm 16.7 (103.22-226.10)	<0.0001
after rewett. (22 d)	^b 181.34 \pm 3.86 (154.95-206.72)	^a 265.44 \pm 3.28 (241.99-287.3)	^a 273.87 \pm 5.24 (232.29-299.92)	^b 144.50 \pm 4.24 (118.30-211.62)	<0.0001
before vs. after-p-value	<0.0001	<0.0001	0.008	<0.0001	

However, a temporal series of the fluxes can help to better distinguish the temporal variation of the present dataset. Figure 3-3 shows the evolution of CO₂ efflux across the 4 treatments, which coincides with the data described in Table 3-2 in the emission ranking of the treatments. Furthermore, it shows high intraday variability (spatial probably), and a strong reaction to the irrigation, mainly on the first day (August 25th) and by the S+N treatment, while the S treatment shows a softer reaction.

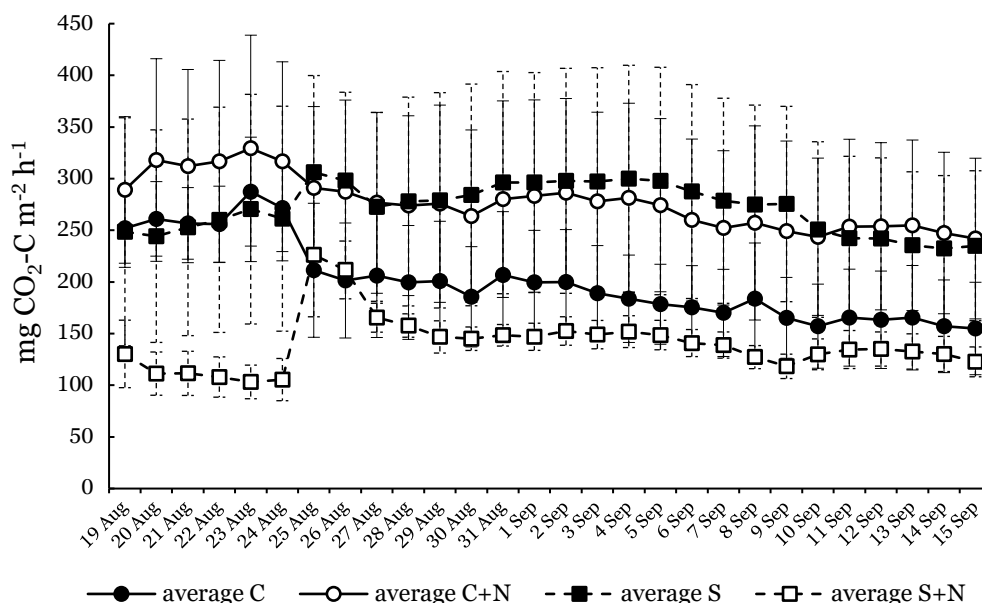


Figure 3-3. Time series of daily averaged soil CO₂ emission [mg CO₂-C m⁻² h⁻¹], per treatment and including nitrogen addition levels, for the 28 days between August 19th and September 15th. Error bars indicate SE.

Similarly, Figure 3-4 also shows the evolution of soil CO₂ efflux in the entire period. However, the data is pooled by precipitation exclusion treatment and averaged, and the new feature here is the different behaviour of the DRW stress plots before and after the rewetting. Also, the average soil water content, natural precipitation, and the amount irrigated are shown. Both soil water content and the CO₂ efflux from the stressed soil react to the irrigation. However, the soil water content in the control plots was more sensitive to water inputs, which is also better and more detailed shown in Figure 2-3 and Figure 2-4. Lastly, the flux curves, mainly control, seem to be coupled with the soil water content.

The fluxes were, on average lower during the 28-d-period ($P<0.01$) and before the rewetting ($P<0.01$) for the stressed plots. Also, the control fluxes were higher before ($P<0.05$) the rewetting. Most importantly, there was no significant difference after the rewetting despite lower soil moisture at the stressed plots (Table 3-3).

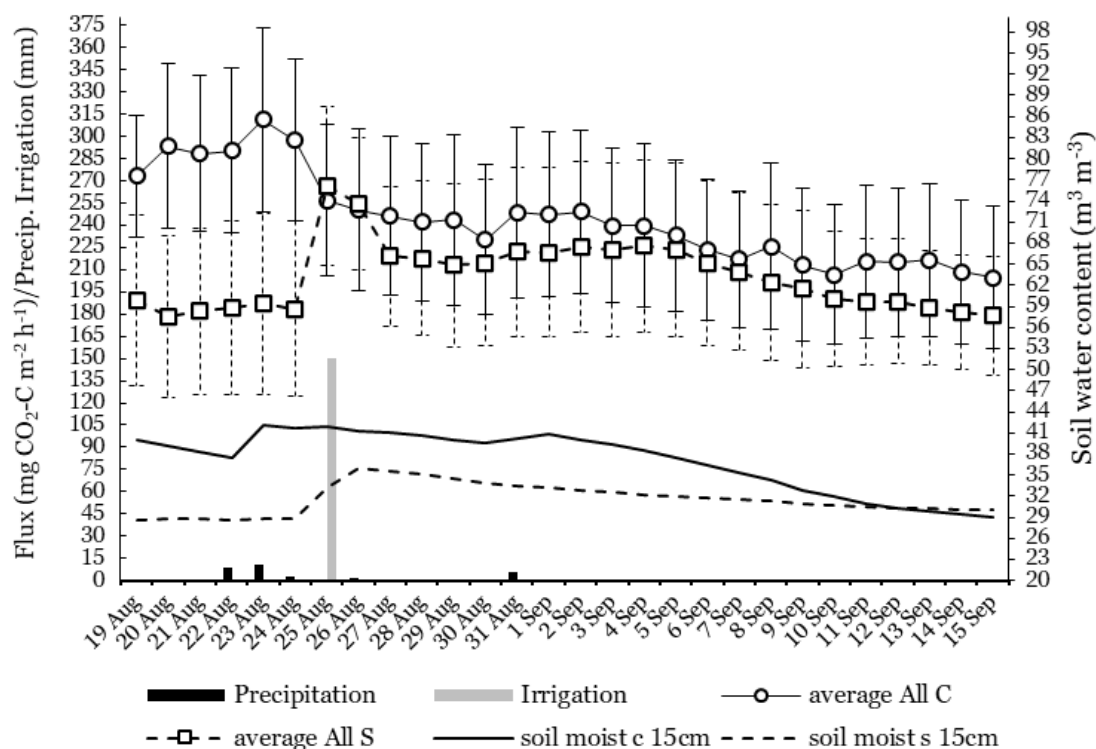


Figure 3-4. Time series of daily averaged soil CO₂ emission [mg CO₂-C m⁻² h⁻¹] (left axis) and soil water content [m³ m⁻³] (right axis), and precipitation/irrigation [mm] (left axis) for the 28 days between August 19th and September 15th, and considering only the rainfall manipulation (pooled and averaged into Average All S and C). Error bars indicate SE.

Table 3-3. Mean \pm SE (min-max) of averaged CO₂ fluxes [mg CO₂-C m⁻² h⁻¹] along days of the two levels of the rainfall manipulation. Three periods were considered, complete (28 days from August 19th to September 15th), before rewetting (6 days from August 19th to 24th), and after rewetting (22 days from August 25th to September 15th). The p-values inform Welch's test results.

	mg CO ₂ -C m ⁻² h ⁻¹		p-value
	control	DRW stress	
complete (28 d)	243.90 \pm 5.69 (204.7-311.3)	205.80 \pm 4.33 (177.8-266.1)	<u>0.008</u>
before rewett. (6 d)	290.50 \pm 5.08 (273.1-311.3)	183.46 \pm 1.63 (177.8-189.1)	<u>0.002</u>
after rewett. (22 d)	245.60 \pm 3.51 (204.7-256.9)	211.78 \pm 4.75 (178.9-266.1)	0.2
before vs after p-value	<u>0.028</u>	0.065	

3.3. Meteorological data

The data related to soil water content and temperature, both differentiated only by precipitation exclusion treatment, and data on precipitation and air temperature were already shown in Figure 2-3, Figure 2-4, Figure 2-5, and Figure 3-4. The means of this data were also tested for

exploring differences. The soil temperature (at 5 cm depth) was similar between treatments but showed a decrease ($P<0.0001$) from the beginning to the end of the analysis period (Table 3-4).

Table 3-4. Mean \pm SE (min-max) of the daily mean soil temperature at 5 cm depth [$^{\circ}\text{C}$] for “control” and “drought” plots averaged and differentiated by period. Three periods were considered, complete (28 days from August 19th to September 15th), before rewetting (6 days from August 19th to 24th), and after rewetting (22 days from August 25th to September 15th). The p-values inform Student’s t-test results.

	Soil temperature 5 cm ($^{\circ}\text{C}$)		p-value
	control	DRW stress	
complete (28 d)	14.54 \pm 0.19 (13.21-16.42)	14.78 \pm 0.19 (13.52-16.66)	0.38
before rewett. (6 d)	15.53 \pm 0.19 (15.21-16.42)	16.18 \pm 0.16 (15.68-16.66)	0.18
after rewett. (22 d)	14.19 \pm 0.17 (13.21-15.76)	14.40 \pm 0.16 (13.52-15.96)	0.37
before vs after p-value	<u>≤ 0.0001</u>	<u>≤ 0.0001</u>	

Figure 2-3 allocates the current thesis analysis period at a moment of maximum difference in soil water content between the treatments. It also shows the already mentioned higher sensitivity to reaction for the control plots. Figure 2-4 zooms in and allows us to see how an irrigation event of 150 mm had the same positive effect as the control plots showed for natural precipitation of ~ 20 mm, which will be discussed later. Lastly, Table 3-5 summarizes the results of the Student’s t-tests, showing higher moisture contents ($P<0.0001$) for the control plots during all the periods analysed and was also higher before the rewetting ($P<0.05$). The DRW stress also shows an increase ($P<0.0001$), but it is being dissolved by the longer 22 days period chosen, so the increase is better seen in the mentioned figures.

Table 3-5. Mean \pm SE (min-max) of daily mean soil water content at 15 cm depth [%] for “control” and “drought” plots averaged and differentiated by period. Three periods were considered, complete (28 days from August 19th to September 15th), before rewetting (6 days from August 19th to 24th), and after rewetting (22 days from August 25th to September 15th). The p-values inform Welch’s t-test results.

	Soil moisture 15 cm (%)		p-value
	control	DRW stress	
complete (28 d)	37.54 \pm 0.67 (30.98-42.18)	28.23 \pm 0.28 (26.10-30.91)	<u>≤ 0.0001</u>
before rewett. (6 d)	39.82 \pm 0.76 (37.48-42.18)	28.07 \pm 0.03 (26.10-26.31)	<u>≤ 0.0001</u>
after rewett. (22 d)	36.92 \pm 0.78 (30.98-41.88)	28.78 \pm 0.25 (27.10-30.91)	<u>≤ 0.0001</u>
before vs after p-value	<u>0.017</u>	<u>≤ 0.0001</u>	

3.4. Regression analysis

Table 3-6 shows the simple and multiple regression models that were significant and showed higher R^2 values. Daily averages of CO_2 fluxes strongly correlated with soil water content and

temperature, in that order in both “control” and “DRW stress” treatments. Furthermore, the multiple regression model relating the CO₂ efflux from control treatment with the soil moisture and temperature as first and second regressor variables showed an R²-value of 0.95, with a positive correlation with soil moisture and temperature. Besides, another linear regression fitted for the DRW stress treatment resulted in soil moisture as a single regressor explaining 66% of the variability on the emission, with a slope value of 12.41. The same model was fitted for the control treatment, and the soil moisture explained 54% of the variation, with a slope of 6.2. On the other hand, soil temperature contrasted in behaviour inside treatments since it correlated positively in “control” but negatively in “DRW stress”. Similar values (R² and slope) were obtained using another approach, with the automatic curve fitting function in Microsoft Excel (Figure 0-1 and Figure 0-2, Appendix A).

Table 3-6. Simple and multiple regression analysis between mean daily CO₂ fluxes against soil temperature and moisture. Pearson’s R² was informed. Significance: * p<0.05, ** p<0.01, *** p<0.001.

Treatment Dependent variable <i>y_t</i>	Rainfall manipulation treatment							
	Control		Control		Drought		Drought	
	mg CO ₂ -C m ⁻² h ⁻¹							
Intercept β ₀	11.1		-373.1	***	-144.52	**	450.16	***
Regression coefficients:								
(soil water content) β _i	6.2	***	8.3	***	12.41	***	--	
(soil temperature) β _i	--		21	***	--		-16.53	***
Total R²:	0.54		0.95		0.66		0.52	
<i>n</i>	28		28		28		28	

3.5. Contour plots

Contour plots are another graphical alternative to represent the relation between variables, like emission, soil temperature, and soil moisture. Figure 3-5 and Figure 3-6 need to be analyzed separately, but in general, the higher the soil temperature and water content, the darker the colour, the higher the soil CO₂ efflux (darker colour). Figure 3-7, which mixes all the values together, has two clouds at mid-low temperature and moisture values, probably corresponding to the pulse after rewetting in the DRW stress treatment.

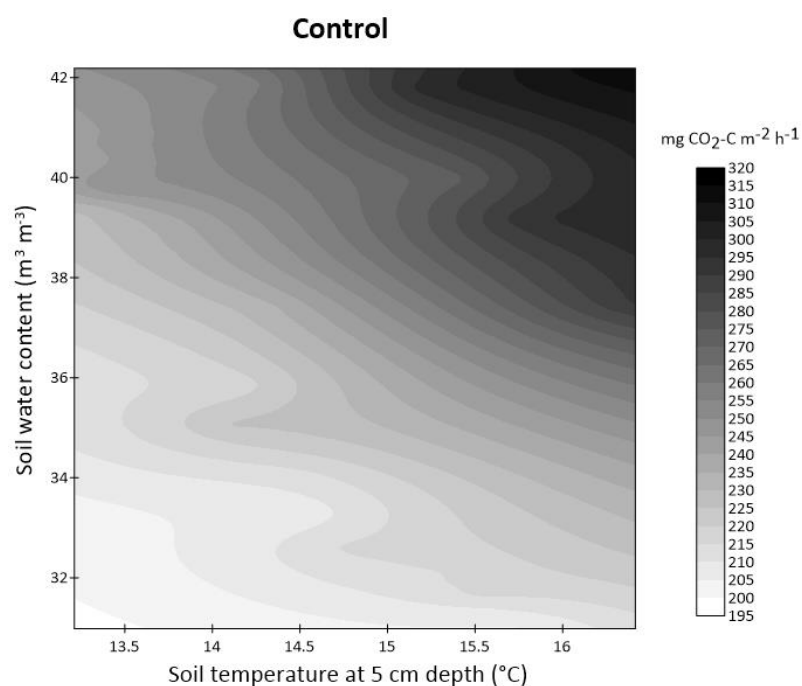


Figure 3-5. Contour map of soil fluxes driven by soil temperature and soil water content in the control treatment. Elaborated with Surfer® (Golden Software, LLC).

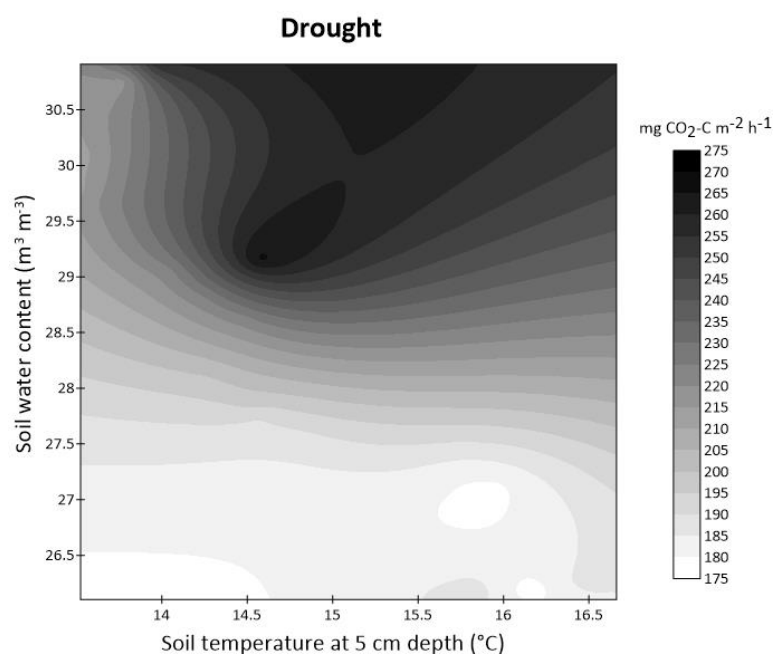


Figure 3-6. Contour map of soil fluxes driven by soil temperature and soil water content in drought treatment. Elaborated with Surfer® (Golden Software, LLC). “Drought” stands for “DRW stress” treatment.

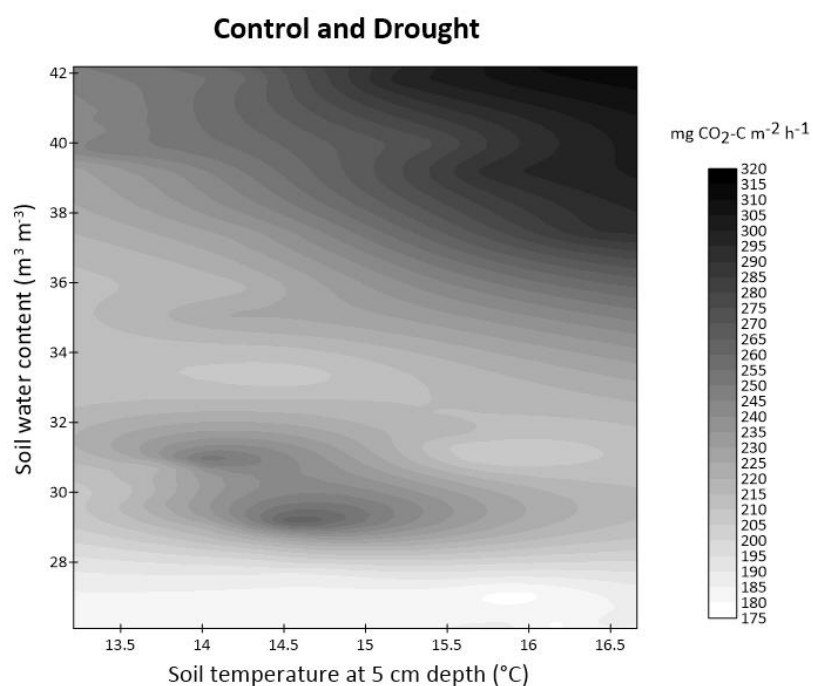


Figure 3-7. Contour map of soil fluxes driven by soil temperature and soil water content in all observations, control and drought treatments combined. Elaborated with Surfer® (Golden Software, LLC). “Drought” stands for “DRW stress” treatment.

4. Discussion

The first hypothesis assumed that the rewet of severely dry soil is expected to reactivate the microbial activity, and so soil CO₂ efflux and MBC would raise after irrigation. The results presented for the temporal variability (section 3.2), including the time series presented in Figure 3-3 and Figure 3-4, which show a strong increase in S and DRW emission on date August 25th, and the significant statistical tests in Table 3-2, support an increased CO₂ efflux after rewetting up to the level of the control. Furthermore, although MBC did not significantly increase after rewetting, it was found a significant correlation between soil CO₂ efflux and MBC at 10 ($\rho = 0.55$; $P < 0.01$) and 20 ($\rho = 0.4$; $P < 0.05$) cm depth and also a linear regression model was fitted ($R^2 = 0.16$; $P < 0.05$; Table 3-1). Likewise, a positive correlation ($R^2 = 0.52$) between soil CO₂ and MBC was also found by Lee & Jose (2003) in a temperate ecosystem. Also, Navarro-García et al. (2011), in a manipulative experiment to assess the effect of aggregation in controlling the microbial response during DRW cycles, found that aggregate destruction, which can also be promoted by DRW cycles, can release occluded C, which can subsequently enhance both Rs and microbial biomass. In the same line, Zhao et al. (2010) also found that increased frequency of DRW cycles, and fertilization events, enhanced not only MBC but also DOC. However, in an incubation study by Sawada et al. (2017), although they found a pulse of Rs due after the DRW cycle, they also described a reduction of MBC whose magnitude relied on soil properties and the history of DRW cycles, where the microbial community in soils with more frequent DRW are better adapted, and so the MBC can be maintained (also demonstrated by Leizaga et al., 2021). These results agree with the current study, where there was no detectable change in the MBC pool. Furthermore, it would be important to monitor what happens in the posterior DRW cycles at Klausen-Leopoldsdorf, to explore if there is really a moderation effect with the increasing history of DRW stress.

Many of the works cited in the previous paragraph did find a pulse after rewetting, and this is the topic of the next hypothesis. The Birch effect can be identified when there is a decoupling of the increasing rate in soil moisture and the Rs pulse, meaning that there is an exacerbated positive response upon an amount of water added over severely dry soil, which does not cause the same Rs pulse in continually moist soil. According to the results also deployed in section 3.2 (“Temporal variability”), there might be some evidence supporting there was a Birch effect pulse type in Rs in the DRW stress treatment. One hint is the sharp increase in S+N treatment shown in Figure 3-3. Also, Table 3-2 shows that although both C and C+N emissions declined in the second period starting on August 25th, at the same time, both S and S+N increased. Likewise, Table 3-3 showed no significant difference between the two pooled treatments (control and DRW stress), and DRW emission might have even surpassed the control level (Figure 3-4). However, the soil in DRW plots have much less water content (Figure 2-4 and Figure 3-4), which is not justified by differences in texture (Table 0-1, Appendix A) and probably neither in structure. Hence, this supports the hypothesis that there was a decoupling between Rs and soil moisture. Also, a plot of the ratio DRW:Control efflux against the ratio of soil moisture (soil moisture in DRW over soil moisture in Control) shows an abrupt jump around a value of 0.7 of the “ratio moisture s/c”, which can support the decoupling (see **Figure 0-3, Appendix A**). Furthermore, a moderate increase in water availability accompanying a

moderate Birch is supported by previous studies. For example, Barnard et al. (2020) proposed the magnitude of the rewetting upshock controls the rate of availability of labile C that feeds the pulse and that upon a certain threshold, occlude organic matter becomes accessible, which was probably not the case at Klausen-Leopoldsdorf in the current DRW cycle, but there was a detection of increased NH_4^+ upon rewetting ($P < 0.05$; Table 0-1), possibly suggesting increased mineralization of organic N, and microbial death can be a source of released C and N labile forms after rewetting (Blazewicz et al., 2014).

A decoupling of the Rs from either the soil moisture and/or the soil temperature is not the rule, but rather the exception in the soil. The third hypothesis assumes these two variables are strong drivers of the Rs. However, the soil moisture can be limiting either by scarcity or abundance, becoming a stronger driver than temperature (Borken et al., 1999; Epron et al., 2004; Yuste et al., 2005), and since there was water limitation (manipulation objective), it can be expected a strong positive correlation between Rs and soil moisture. The regression models in Table 3-6 support the idea that both variables were important drivers, even though maybe it was not expected from the temperature. However, soil moisture was stronger as a predictor and had a positive relation. Similar results have been found by Kitzler et al. (2006) at the same site. Notwithstanding, what was maybe not expected was the high variability not explained by moisture and temperature that can be attributed to spatial variability among plots and will be discussed in the next hypothesis. What also remains elusive is the fate of the irrigation water. The soil water content in the control plots was much more sensitive to inputs since it can be seen in its pulses to natural precipitation events, e. g. of a magnitude ~20 mm in two consecutive days (August 22nd and 23rd; Figure 2-4). This event generated an absolute increase of ~5% in the soil water content, while by the irrigation event of 150 mm, the DRW plots increased ~5%. This would be interesting to explore further. One possible explanation would be that the rate of precipitation surpasses the capacity of the soil to retain the water, which loses it by deep percolation or superficial sub-superficial runoff. In line with this, drought can promote hydrophobicity (Goulden et al., 1996), which would enhance superficial runoff that was not detected at the moment of irrigation. However, a superficial runoff was not detected in the field.

There were several hints pointing at the existence of a high variability among plots in the current setup. We hypothesized that since forest soils show high variability of soil biochemical and physical parameters, CO_2 efflux would also show high spatial variability, and it would be possible to find correlations with any of the biochemical and physical soil parameters monitored. The importance of spatial variability among plots in the current study can be depicted by comparing Figure 3-1 vs Figure 3-2. While the first one only includes spatial variability because it is reduced to only two days, the second one includes the 28 measurements of the entire period analysed. Surprisingly, they are very similar, meaning that the added temporal variability might not be so important. Also, the time series plots Figure 3-3 and Figure 3-4 also denote a huge variability at every daily average, represented by the error bars (error standard). The explanatory power over Rs of the spatial variability has been studied not only in forests soils but also in general when studying trace gas flux emissions (Bahn et al., 2008; Curiel Yuste et al., 2004; Epron et al., 2004b; Kang et al., 2003; Law et al., 2001; Sørensen & Buchmann, 2005; Vincent et al., 2006). For example, Sørensen & Buchmann, 2005 working on a beech forest in Germany, developed heat maps of Rs and correlated them with several soil and

vegetation parameters. This allows them to identify “hotspots” of emission. In the current setup, we might be able to also find them. For example, the high emission of plots 2 and 3 might correspond with hotspots. Here, in particular, it might be important to check if autotrophic (plant roots and rhizospheric) respiration is inflating the emission. With the purpose of disentangling the spatial patterns of Rs in the current setup, the correlations and regression shown in Table 3-1 were performed. The exploration resulted in the identification of a few parameters with some potential to explain Rs spatial heterogeneity. Those were Total carbon at 10 cm (log-transformed), MBC at 10 cm, MBN at 10 cm, and DOC at 10 cm, which along with other variables (e.g. pH, Bulk density, Litter layer depth), were also found to explain Rs in others works (Bahn et al., 2008; Curiel Yuste et al., 2004; Epron et al., 2004b; Kang et al., 2003; Law et al., 2001; Sørensen & Buchmann, 2005; Vincent et al., 2006). Assessing spatial variability would be essential to future model attempts of Rs.

Finally, another expectation was that enrichment of forest soils with Nr would increase the concentration of mineral forms of N, either NO_3^- or NH_4^+ , reduce microbial activity (MBC), and suppress soil CO_2 efflux. Among the results, the only significant difference found when comparing the N addition treatment was in C:N ratio at 20 cm. However, even this difference might not have been caused by the N treatment since it gave the opposite results, showing lower values (higher N content) for the untreated soil. Furthermore, there was a not expected pattern in the Rs (Table 3-2), while Rs might have been enhanced by the N addition in the control plots, there was a suppression effect in the DRW stress treatment. The latter effect might be explained by the combination of the expected depression effect that Nr has been described to have in the Rs (Dirnböck et al., 2017; Janssens et al., 2010; Pregitzer et al., 2008; Zhou et al., 2014; Zhong et al., 2016), and the suppression by DRW stress which can also hamper the Rs. However, further exploration might shed light on whether this apparent enhancing effect of the N addition on the control plots is reproducible or if it is a product of spatial variability.

5. Conclusions

The present study offers a reliable set of measurements of soil respiration of a mature temperate forest and an effort to disentangle the contribution of soil parameters on the temporal and spatial variability under a simulated scenario of increased dry-rewetting stress and atmospheric Nr deposition.

This thesis found evidence suggesting the occurrence of a moderated Birch effect and a rapid recovery of the soil respiration upon rewetting. Also, this response would be led not only by soil moisture content but by other soil parameters, which enhances the spatial variability of the soil respiration.

Although soil moisture and temperature were found to be the most important drivers of soil respiration, a vast spatial variability has been described, and the current study gives preliminary clues over which soil parameters might be driving that.

Furthermore, this thesis tried to distinguish between the Birch effect and the soil moisture, which usually overlap.

Finally, the analysis of the effect of atmospheric Nr deposition throws a new pattern which has to be confirmed with further exploration.

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Appendix A: Supplementary data

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<i>parameter</i>	Precipitation exclusion			Prec. Exc. + Nitrogen addition				
	control	DRW stress	<i>p-value</i>	c	c + n	s	s + n	<i>p-value</i>
<i>sand [%] (10 cm)</i>	14.22 \pm 1.47 (10.17-23.47)	12.48 \pm 0.93 (9.21-17.07)	0.33	15.76 \pm 2.63 (12.37-23.47)	12.69 \pm 1.26 (10.17-16.19)	13.86 \pm 1.51 (9.91-17.07)	11.11 \pm 0.71 (9.21-12.40)	0.3
<i>sand [%] (20 cm)</i>	11.17 \pm 0.76 (8.59-15.97)	10.64 \pm 0.60 (8.85-13.28)	0.59	12.32 \pm 1.23 (10.86-15.97)	10.03 \pm 0.59 (8.59-11.29)	11.07 \pm 1.12 (8.94-13.28)	10.22 \pm 0.56 (8.85-11.57)	0.33
<i>silt [%] (10 cm)</i>	55.32 \pm 1.62 (51.89-62.69)	55.13 \pm 1.61 (47.22-60.06)	0.93	52.54 \pm 0.22 (51.89-52.80)	58.10 \pm 2.66 (52.04-62.69)	54.34 \pm 1.69 (50.65-57.70)	55.92 \pm 2.96 (47.22-60.06)	0.36
<i>silt [%] (20 cm)</i>	52.96 \pm 0.73 (49.27-56.51)	55.74 \pm 1.15 (51.23-60.00)	0.061	52.01 \pm 1.00 (49.25-53.85)	53.92 \pm 0.95 (51.98-56.51)	55.22 \pm 1.61 (52.66-59.87)	56.26 \pm 1.84 (51.23-60.00)	0.22
<i>clay [%] (10 cm)</i>	30.4 \pm 1.58 (23.73-35.73)	32.39 \pm 1.31 (28.78-40.38)	0.36	31.7 \pm 2.75 (23.73-35.73)	29.21 \pm 1.76 (25.18-32.53)	31.8 \pm 1.30 (28.78-34.43)	32.98 \pm 2.48 (29.88-40.38)	0.66
<i>clay [%] (20 cm)</i>	35.86 \pm 1.04 (30.18-39.78)	33.62 \pm 0.98 (30.78-38.4)	0.14	35.68 \pm 2.00 (30.18-39.78)	36.05 \pm 1.01 (33.88-38.1)	33.71 \pm 1.67 (30.78-38.4)	33.53 \pm 1.29 (31.15-37.2)	0.56
<i>clay [%] (20 cm)</i>	3.27 \pm 0.21 (2.53-4.17)	3.85 \pm 0.31 (2.91-5.56)	0.14	3.14 \pm 0.35 (2.53-3.79)	3.40 \pm 0.27 (3.03-4.17)	4.08 \pm 0.52 (3.16-5.56)	3.63 \pm 0.36 (2.91-4.53)	0.41
<i>Total carbon [w/w%] (20 cm)</i>	2.09 \pm 0.23 (1.26-3.36)	2.70 \pm 0.34 (1.60-4.35)	0.16	2.09 \pm 0.47 (1.26-3.36)	2.09 \pm 0.17 (1.82-2.54)	2.73 \pm 0.41 (1.60-3.46)	2.67 \pm 0.60 (1.64-4.35)	0.6
<i>Total nitrogen [w/w%] (10 cm)</i>	0.17 \pm 0.01 (0.1-0.21)	0.20 \pm 0.02 (0.14-0.26)	0.19	0.16 \pm 0.02 (0.1-0.19)	0.19 \pm 0.02 (0.15-0.21)	0.22 \pm 0.03 (0.14-0.26)	0.19 \pm 0.02 (0.15-0.23)	0.26

	Total nitrogen [w/w%] (20 cm)	0.10 ± 0.01 (0.06-0.17)	0.15 ± 0.02 (0.07-0.22)	<u>0.04</u>	0.1 ± 0.03 (0.06-0.17)	0.1 ± 0.01 (0.08-0.12)	0.17 ± 0.03 (0.1-0.22)	0.13 ± 0.03 (0.07-0.21)	0.16
	C:N 10 cm	19.44 ± 1.36 (14.85-26.26)	19.27 ± 1.00 (15.27-22.38)	0.92	20.81 ± 2.22 (15.50-26.26)	18.06 ± 1.58 (14.85-20.78)	18.84 ± 1.91 (15.27-22.38)	17.70 ± 0.93 (17.60-22.04)	0.71
	C:N 20 cm	21.92 ± 1.16 (17.80-27.33)	18.21 ± 1.07 (15.37-24.50)	<u>0.04</u>	^{ab} 21.56 ± 1.34 (19.52-25.22)	^a 22.28 ± 2.09 (17.80-27.33)	^c 16.10 ± 0.34 (15.37-17.02)	^{bc} 20.33 ± 1.51 (18.12-24.50)	<u>0.04</u>
before rewett.	microbial biomass carbon [C mg (soil kg) ⁻¹] (10 cm)	185.23 ± 14.44 (117.82-233.80)	142.19 ± 15.43 (79.17-209.5)	0.06	174.31 ± 24.04 (117.82- 233.8)	196.16 ± 17.76 (146.38- 227.2)	137.04 ± 30.3 (79.17-209.5)	147.34 ± 13.24 (108.25- 163.71)	0.28
after rewett.		131.55 ± 12.91 (67.21-197.25)	119.61 ± 9.18 (98.35-176.54)	0.79	128.86 ± 26.78 (67.21- 197.25)	134.24 ± 7.50 (120.31- 153.39)	128.03 ± 18.05 (98.35- 176.54)	111.18 ± 4.53 (101.50- 119.80)	0.46
p-value		<u>0.015</u>	0.23		0.25	0.03	0.81	0.066	
before rewett.	microbial biomass carbon [C mg (soil kg) ⁻¹] (20 cm)	167.68 ± 27.76 (86.18-290.4)	170.92 ± 33.13 (62.64-349.13)	0.94	169.17 ± 42.06 (110.22- 290.4)	166.18 ± 42.72 (86.18- 286.68)	161.32 ± 33.54 (98.26- 255.58)	180.52 ± 62.75 (62.64- 349.13)	0.99
after rewett.		116.63 ± 18.20 (65.25-213.94)	123.08 ± 9.74 (88.31-169.72)	0.76	120.53 ± 33.58 (65.25- 213.94)	112.74 ± 20.18 (81.85- 171.23)	130.69 ± 17.11 (94.12- 169.72)	115.47 ± 10.55 (88.31- 136.55)	0.94
p-value		0.15	0.2		0.4	0.32	0.46	0.38	
before rewett.	microbial biomass nitrogen [N mg (soil kg) ⁻¹] (10 cm)	24.36 ± 2.22 (17.0-33.88)	15.44 ± 2.34 (7.64-24.89)	<u>0.015</u>	23.58 ± 3.9 (17.0-33.88)	25.13 ± 2.75 (17.03-29.19)	15.81 ± 3.99 (7.64-24.89)	15.08 ± 3.08 (8.08-20.61)	0.136
after rewett.		18.60 ± 1.31 (11.61-22.72)	18.34 ± 1.72 (12.19-26.11)	0.9	17.67 ± 2.32 (11.61-22.72)	19.53 ± 1.43 (15.30-21.48)	19.57 ± 3.26 (12.19-26.11)	17.11 ± 1.46 (13.43-20.54)	0.81
p-value		<u>0.047</u>	0.34		0.25	0.14	0.49	0.58	
before rewett.	microbial biomass nitrogen [N mg (soil kg) ⁻¹] (20 cm)	22.94 ± 4.25 (11.73-48.15)	19.54 ± 4.31 (5.35-44.58)	0.58	25.1 ± 7.95 (13.36-48.15)	20.78 ± 4.25 (11.73-31.88)	18.0 ± 3.21 (12.79-27.02)	21.06 ± 8.65 (5.35-44.58)	0.89
after rewett.		16.59 ± 1.47 (9.8±23.06)	21.86 ± 2.77 (14.31-37.42)	0.12	17.07 ± 2.81 (9.86-23.06)	16.11 ± 1.41 (13.23-18.95)	24.00 ± 5.26 (14.62-37.42)	19.71 ± 2.28 (14.31-25.42)	0.36
p-value		0.19	0.66		0.4	0.36	0.38	0.89	

<i>before rewett.</i>		197.97 ± 13.83 (135.56-251.45)	216.72 ± 11.83 (184.35-289.62)	0.89	175.68 ± 17.15 (135.56- 219.29)	220.27 ± 16.35 (187.89- 251.45)	203.84 ± 11.83 (184.35- 236.87)	229.6 ± 20.05 (205.77- 289.62)	0.32
	<i>dissolved organic carbon [C mg (soil kg)⁻¹] (10 cm)</i>								
<i>after rewett.</i>		205.32 ± 16.28 (127.55-267.55)	210.86 ± 8.33 (175.64-245.12)	0.77	192.64 ± 27.97 (127.5- 244.5)	217.99 ± 18.62 (181.46- 267.55)	215.31 ± 14.52 (175.64- 245.12)	206.41 ± 10.00 (181.29- 229.20)	0.78
<i>p-value</i>		0.74	0.69		0.63	0.93	0.56	0.35	
<i>before rewett.</i>		175.66 ± 12.65 (133.25-239.36)	206.41 ± 17.2 (150.56-280.73)	0.17	160.78 ± 14.98 (133.25- 201.26)	190.54 ± 19.35 (154.93- 239.36)	176.49 ± 8.83 (150.56- 188.42)	236.33 ± 26.6 (172.5- 280.73)	0.069
	<i>dissolved organic carbon [C mg (soil kg)⁻¹] (20 cm)</i>								
<i>after rewett.</i>		169.47 ± 12.45 (128.01-213.66)	192.33 ± 10.50 (163.83-249.19)	0.18	165.58 ± 21.50 (128.01- 205.16)	173.36 ± 15.79 (147.23- 213.66)	194.77 ± 10.68 (165.13- 215.19)	189.89 ± 19.90 (163.83- 249.19)	0.62
<i>p-value</i>		0.73	0.5		0.86	0.52	0.24	0.22	
<i>before rewett.</i>		23.24 ± 1.95 (14.22-33.1)	25.5 ± 1.42 (19.53-31.47)	0.37	22.71 ± 3.91 (14.22-33.1)	24.78 ± 1.52 (20.08-27.17)	26.68 ± 2.51 (19.53-31.47)	26.31 ± 1.64 (23.15-30.74)	0.79
	<i>dissolved organic nitrogen [N mg (soil kg)⁻¹] (10 cm)</i>								
<i>after rewett.</i>		24.78 ± 1.65 (15.82-30.82)	30.00 ± 2.31 (21.17-40.71)	0.086	23.25 ± 3.18 (15.82-30.82)	26.30 ± 1.00 (24.71-28.94)	31.64 ± 4.08 (21.17-40.71)	28.36 ± 2.51 (23.97-35.36)	0.275
<i>p-value</i>		0.56	0.12		0.92	0.22	0.21	0.52	
<i>before rewett.</i>		21.73 ± 1.14 (15.74-25.54)	24.44 ± 1.78 (17.81-33.64)	0.22	20.64 ± 2.11 (15.74-25.54)	22.81 ± 0.93 (20.7-24.67)	22.15 ± 2.07 (17.81-26.29)	26.73 ± 2.62 (22.33-33.64)	0.23
	<i>dissolved organic nitrogen [N mg (soil kg)⁻¹] (20 cm)</i>								
<i>after rewett.</i>		21.14 ± 1.84 (15.16-30.73)	24.03 ± 1.49 (19.55-31.15)	0.24	19.31 ± 2.48 (15.16-25.93)	22.97 ± 2.72 (18.68-30.73)	24.02 ± 2.08 (19.55-28.89)	24.05 ± 2.46 (20.24-31.15)	0.5
<i>p-value</i>		0.79	0.86		0.7	0.96	0.55	0.49	
<i>before rewett.</i>		3.05 ± 1.18 (0.38-9.87)	3.22 ± 0.62 (1.10-5.95)	0.9	3.00 ± 2.30 (0.38-9.87)	3.10 ± 1.08 (1.52-6.15)	1.85 ± 0.44 (1.10-3.13)	4.59 ± 0.60 (3.13-5.95)	0.56
	<i>nitrate [μg NO₃-N (g dry weight)⁻¹ (10 cm)]</i>								
<i>after rewett.</i>		1.59 ± 0.46 (0.11-3.68)	2.55 ± 0.63 (0.11-5.59)	0.23	1.58 ± 0.53 (0.55-2.88)	1.60 ± 0.83 (0.11-3.68)	2.22 ± 0.53 (0.86-3.45)	2.91 ± 1.22 (0.11-5.59)	0.64
<i>p-value</i>		0.28	0.47		0.59	0.31	0.61	0.28	

<i>before rewett.</i>	<i>nitrate [$\mu\text{g NO}_3\text{-N}$ (g dry weight)⁻¹] (20 cm)</i>	1.67 ± 0.34 (0.56-3.52)	1.68 ± 0.32 (0.76-3.54)	0.98	1.52 ± 0.38 (0.86-2.54)	1.82 ± 0.62 (0.56-3.52)	1.28 ± 0.18 (0.87-1.68)	2.09 ± 0.59 (0.76-3.54)	0.66
<i>after rewett.</i>		1.36 ± 0.31 (0.22-3.33)	1.08 ± 0.12 (0.63-1.77)	0.42	0.89 ± 0.24 (0.22-1.22)	1.84 ± 0.50 (1.22-3.33)	1.33 ± 0.15 (1.16-1.77)	0.84 ± 0.08 (0.63-0.99)	0.1
<i>p-value</i>		0.52	0.12		0.22	0.98	0.83	0.12	
<i>before rewett.</i>	<i>ammonium [$\mu\text{g NH}_4\text{-N}$ (g dry weight)⁻¹] (10 cm)</i>	7.05 ± 1.31 (3.27-15.71)	7.04 ± 0.51 (4.51-8.92)	0.99	8.48 ± 2.45 (4.94-15.71)	5.63 ± 0.83 (3.27-7.02)	7.20 ± 0.37 (6.26-7.87)	6.88 ± 1.04 (4.51-8.92)	0.57
<i>after rewett.</i>		5.93 ± 0.85 (3.21-11.07)	8.94 ± 1.11 (5.07-15.17)	<u>0.0495</u>	5.08 ± 0.76 (3.21-6.40)	6.78 ± 1.53 (4.26-11.07)	10.08 ± 1.70 (8.13-15.17)	7.80 ± 1.42 (5.07-11.76)	0.14
<i>p-value</i>		0.49	0.15		0.26	0.54	0.16	0.62	
<i>before rewett.</i>	<i>ammonium [$\mu\text{g NH}_4\text{-N}$ (g dry weight)⁻¹] (20 cm)</i>	6.14 ± 0.53 (3.90-8.68)	5.76 ± 0.46 (4.16-7.19)	0.59	6.28 ± 0.99 (3.90-8.68)	6.01 ± 0.56 (4.92-7.49)	5.58 ± 0.71 (4.16-7.06)	5.93 ± 0.68 (4.18-7.19)	0.93
<i>after rewett.</i>		5.38 ± 0.98 (2.23-11.47)	5.28 ± 0.42 (3.83-7.29)	0.93	3.97 ± 0.65 (2.23-5.19)	6.79 ± 1.66 (3.68-11.47)	5.38 ± 0.50 (4.03-6.32)	5.17 ± 0.75 (3.83-7.29)	0.31
<i>p-value</i>		0.51	0.46		0.11	0.68	0.83	0.48	

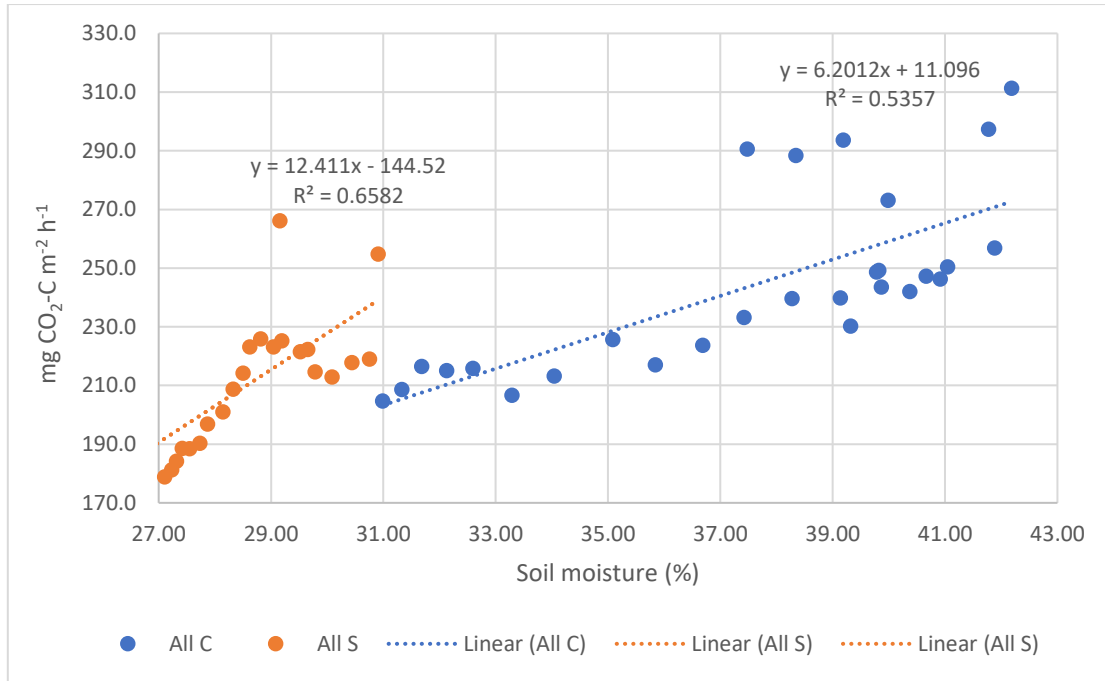


Figure 0-1. Scatterplot of soil efflux ($\text{mg CO}_2\text{-C m}^{-2} \text{ h}^{-1}$) against soil moisture. "All C" refers to averaged in control plots while "All S" corresponds to "DRW stress". The dashed line results from the automated curve fitting function in Microsoft Excel.

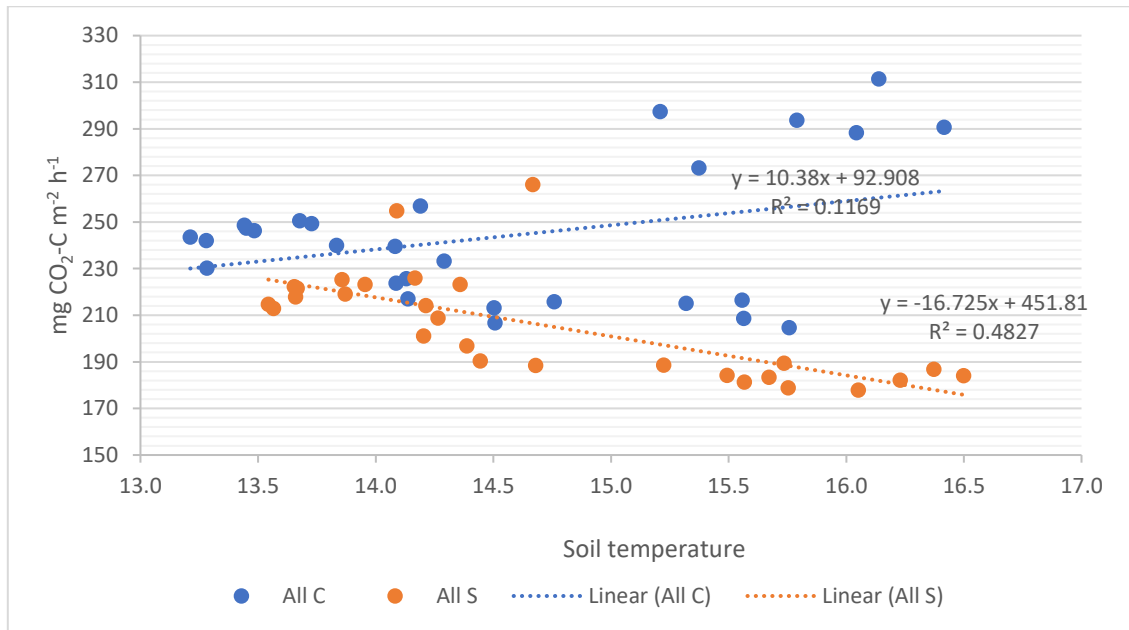


Figure 0-2. Scatterplot of soil efflux ($\text{mg CO}_2\text{-C m}^{-2} \text{ h}^{-1}$) against soil temperature. "All C" refers to averaged in control plots while "All S" corresponds to "DRW stress". The dashed line resulted from the automated curve fitting function in Microsoft Excel.

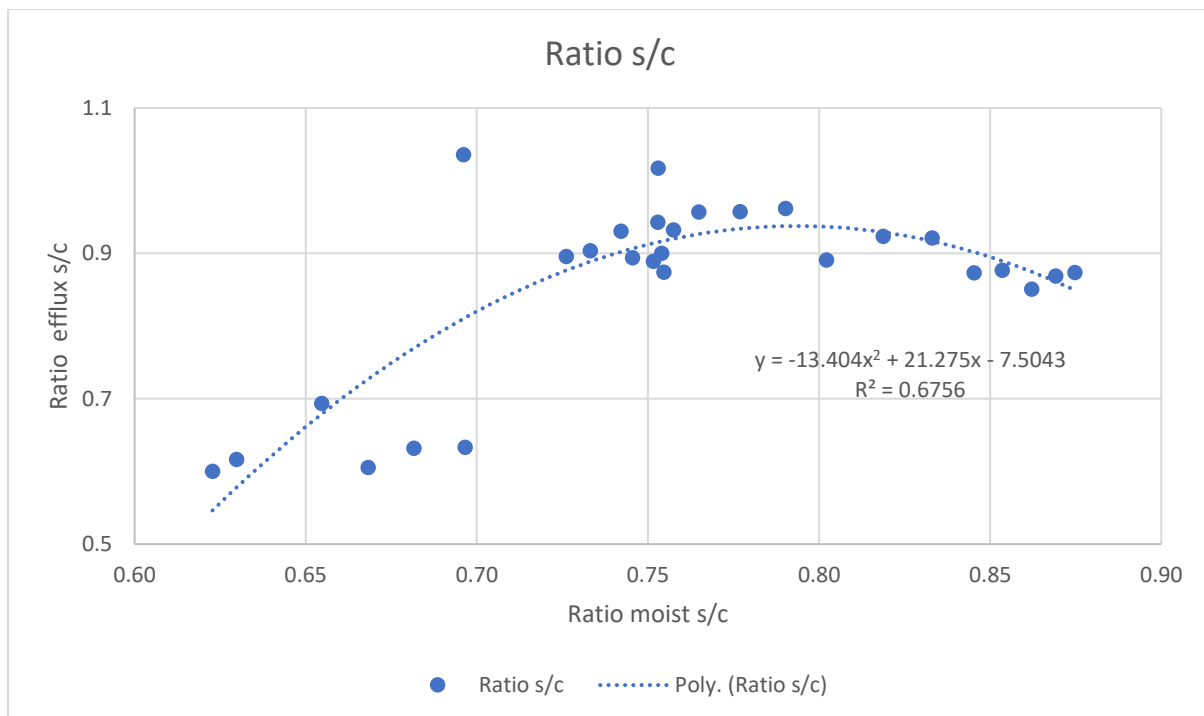


Figure 0-3. Ratio soil CO₂ efflux DRW:Control (ratio efflux s/c) plotted against the ratio soil moisture DRW:Control (ratio moist s/c). There is an abrupt change in ratio efflux s/c around a value of ratio moist s/c around 0.7, which can indicate a decoupling between soil CO₂ efflux and the soil moisture content. Idea extracted from other work (missing reference) and adapted to the current thesis data. A polinomial curve was fitted used Microsoft Excel curve estimation.